

P3-09. DUPLICATED GENES UNDER CONCERTED EVOLUTION IN *Helicobacter*

pylori

Jerson Alexander García-Zea, Rafael Navajas-Pérez, Francisca Robles, Juan Felipe Zapata, Roberto de la Herrán, Carmelo Ruiz Rejón

Department of Genetics, University of Granada, Spain, Faculty of Forensic Science, Technology of Antioquia, Colombia. Corresponding Author: Jerson Alexander García-Zea (alexander7719@correo.ugr.es)

Helicobacter pylori, a Gram-negative bacillus, is present in more than half of the world population. Colonizes the human stomach inducing the development of superficial gastritis, gastric adenocarcinoma, and mucosa-associated lymphoid tissue lymphoma.

H. pylori is highly variable thanks to a great genomic dynamic promoted by mutations and high recombination rates. Also characterized by exhibiting panmixia, genetic mosaicism, and great genomic plasticity, which indicates its ability to adapt at a micro-evolutionary level to its environment (1).

This dynamic influences in the gene duplication events and in the evolution of them. Concerted evolution (CE) between duplicate genes can occur through a genetic exchange called gene conversion (2). Few studies have analyzed the CE and the affected genes in this bacterium (3). Implementing a computational framework (including IseeCe program) (4) in 53 complete *H. pylori* genomes of five different geographical origins, 8 genes (*HopJ/Hopk*, 3'-5' exonuclease, fucosyltransferase, glycosyltransferase, *oipA*, RMs, 50S ribosome and *cagA*) were detected showing patterns of CE. The gene *HopJ/Hopk* shows CE in the five geographical regions, 3'-5' exonuclease, fucosyltransferase in Asia and Amerindian, glycosyltransferase, *oipA*, RMs in populations of Asia only, and finally 50S ribosome and *cagA* showed CE in hybrid strains. Different test by RDP5 software evidence recombination events in the majority of the genes analyzed.

Additionally, we realized analysis of selective pressures on the groups of genes under CE by calculating the non-synonym/synonym mutations ratio (dN/dS) over the complete sequences. Proteins of these genes play crucial roles in the pathogenic life cycle of *H. pylori* such as virulence, pathogenicity, colonization of the environment, adaptation to the host.

1. Garcia-Zea et al. (2019) PeerJ 11,7:e6221
2. Wang S et al. (2018) Commun Biol. 8, 1-12
3. Pride DT et al. (2020) J Mol Biol. 316, 629-42
4. Meinersmann RJ, et al. (2010) Lett Appl Microbiol. 51, 539-545



Facultad de Ciencias

DUPLICATED GENES UNDER CONCERTED EVOLUTION IN *Helicobacter pylori*

Jerson Alexander García-Zea, Rafael Navajas-Pérez, Francisca Robles, Juan Felipe Zapata,

Roberto de la Herrán, Carmelo Ruiz Rejón

Department of Genetics, University of Granada, Spain,

Faculty of Forensic Science, Technology of Antioquia, Colombia

Corresponding author: Jerson Alexander García-Zea (alexander7719@correo.ugr.es)

BACKGROUND

Helicobacter pylori, a gram-negative bacillus, is present in more than half of the world's population. It colonizes the human stomach, inducing the development of superficial gastritis, gastric adenocarcinoma, and mucosa-associated lymphoid tissue lymphoma. *H. pylori* is highly variable thanks to great genomic dynamics promoted by mutations and high rates of recombination. It is also characterized by exhibiting pannikinia, genetic mosaicism and great genomic plasticity, which indicates its ability to adapt at a microevolutionary level to its environment (1). This dynamics influences gene duplication events and their evolution, as is already evident in the genes that encode the membrane proteins *babA* / *babB*, and *HomA* / *HomB* in *H. pylori*. Concerted evolution (CE) between duplicate genes can occur through a genetic exchange called gene conversion (2). Few studies have analyzed EC and affected genes in this bacterium (3).

MATERIALS AND METHODS

A computational framework that included *IseeCe* was implemented to detect CE for gene conversion (GCU) based on phylogeny and genomic synthesis (Mauve) in 53 complete *H. pylori* genomes, from five different geographic origins. Recombination was studied by RDP5 (RDP, Maxchi, Genecon, Bootscan, Chimera, Sliscan, 3seq, Lard and Phylopro). Syntonymous / non-syntonymous substitutions was calculated by ETE3 evol-PAML (models M0, M1, M2, M3, M7 and M8) and natural selection by MegaX (Tajima's tests of neutrality)

RESULTS and DISCUSSION

In total, eight genes were identified in 16 clusters that exhibited CE patterns in different geographical regions, belonging all of them to variable genome, except 50S ribosome-binding GTPase which is part of the central genome. The HopJ / HopK gene exhibited CE patterns in all five geographic groups (Africa, Europe, Asia, Amerindians, and hybrids). In the Amerindian strains, a duplication of the cluster HopJ / HopK can be observed (Table 1). The fucosyltransferase gene showed CE in the Asian and Amerindian groups. The Asian, Amerindian and hybrid strains, 3'-5' exonuclease, oipA, and restriction endonuclease (hsdS) only showed CE for the genes for glycosyltransferase, oipA, and restriction endonuclease (hsdS), only showed CE for the specific Asian group and the oncogenic effector gene of the type IV secretion system (cagA) only in Amerindians. Finally, GTPase gene that binds to the 50S ribosome shows EC in the hybrid group (Table 1).

Comparative sequence analysis provided evidence of recombination between paralogs in 13 of the 16 clusters, evidence of balancing selection in 4 clusters for the gene hopJ/HopK and in one cluster for the gene alpha-(1,3)-fucosyltransferase and evidence of selective sweep in the rest of the clusters, supporting concerted evolution result (Table 1). Both mechanisms stimulate the homogenization of sequences and the functionality of the paralogous gene. In the case of duplicate cluster, hopJ/HopK in Amerind region, we can observe a different identity between clusters in the paralogous genes, being the more identity (95%) under selection sweep and the other cluster with low identity (5%) under balancing selection. This difference in this duplicate cluster could indicate a pseudogenization process in the latter cluster.

Figure 1. CE for 195 HopJ / HopK detected from *IseeCe*. In red duplicate genes that have evolved under CE evolution.

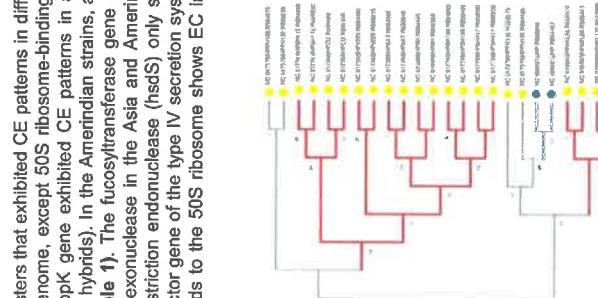


Table 1. Statistical summary. %: Percentage of substitutions, ds / dn: synonymous substitutions and non-synonymous substitutions, Tajima's Test: D = D (neutral equilibrium), D > 0 (balancing selection), D < 0 (recent selective sweep), R = recombination.

Gene	Tajima's Neutrality Test			
	1%	ds/dn	R	
1_15_glycosyltransferase (Asia)	75	0.40492	Recent selective sweep	
2_107_SOS ribosome-binding GTPase (Hybrids)	89	0.47053	Recent selective sweep	
3_90_restriction endonuclease subunit S (Asia)	49	1.48838	Recent selective sweep	
4_30_type IV secretion system (Amerind)	81	1.38924	Recent selective sweep	
5_54_outer inflammatory protein OppA (Asia)	98	0.23353	Recent selective sweep	
6_68_3'-5' exonuclease (Asia)	96	0.16825	Recent selective Not	
7_124_3'-5' exonuclease (Amerind)	97	0.16694	Recent selective Not	
8_88_alpha-(1,2)-fucosyltransferase (Asia)	82	0.22449	Recent selective sweep	
9_89_alpha-(1,3)-fucosyltransferase (Amerind)	88	0.93992	Recent selective sweep	
10_106_alpha-(1,3)-fucosyltransferase (Hybrids)	86	0.27713	Recent selective sweep	
11_136_hopJ/HopK (Africa)	92	0.17239	Balancing selection	
12_177_hopJ/HopK (Europe)	94	0.23891	0.048842	Balancing selection
13_152_hopJ/HopK (Asia)	94	0.33026	0.053488	Balancing selection
14_156_hopJ/HopK (Amerind)	54	0.49336	0.088041	Balancing selection
15_195_hopJ/HopK (Amerind)	95	0.25624	0.048580	Recent selective sweep
16_134_hopJ/HopK (Hybrids)	92	0.24218	0.064525	0.076667 0.847879 Balancing selection

- REFERENCES:
1. García-Zea et al. (2019) PeerJ 11:e62221
 2. Wang S et al. (2018) Commun Biol. 8: 1-12
 3. Pride DT et al. (2020) J Mol Biol. 316, 629-42
 4. Meinersmann RJ et al. (2010) Lett Appl Microbiol. 51, 539-545

D = D (neutral equilibrium), D > 0 (balancing selection), D < 0 (recent selective sweep), R = recombination.