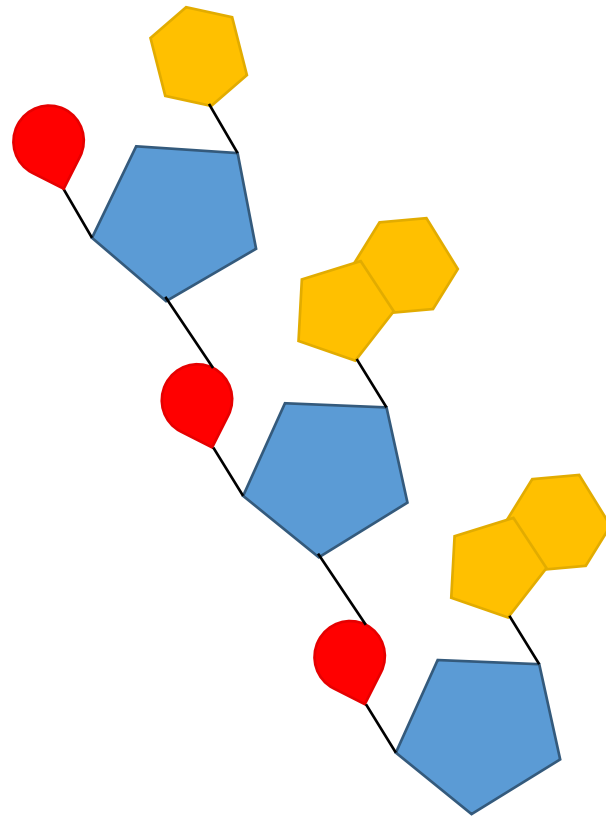
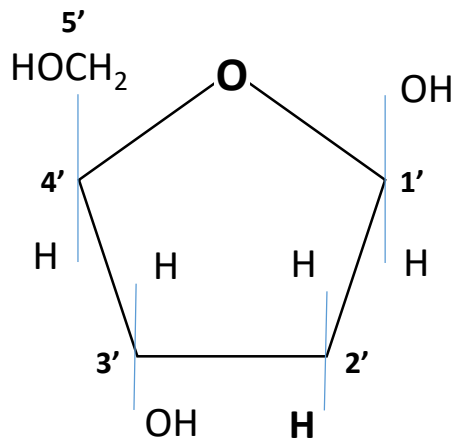


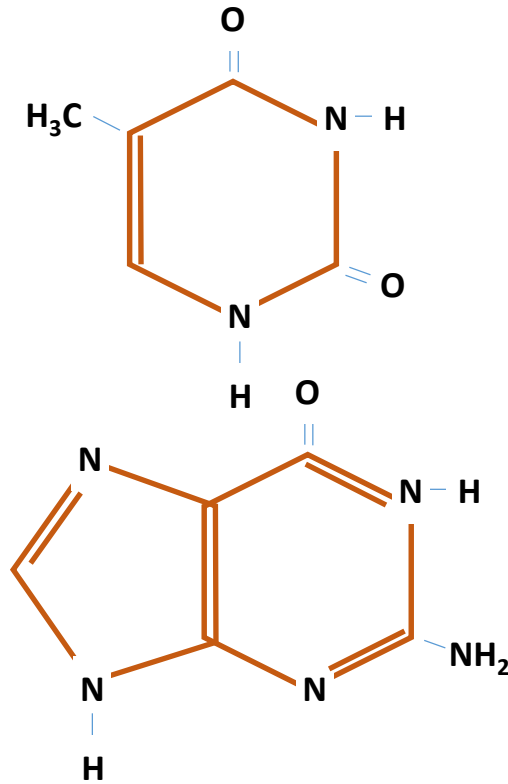
Nature, structure and spatial organization of hereditary material



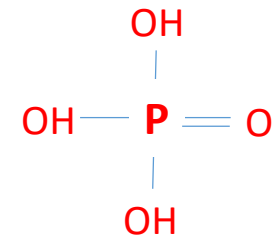
DNA composition



Pentose
(Deoxyribose)

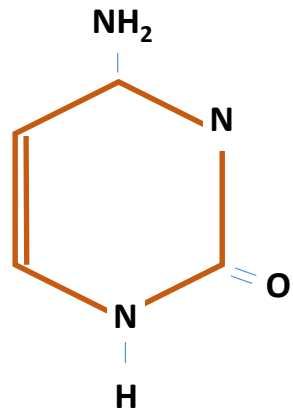


Nucleobase

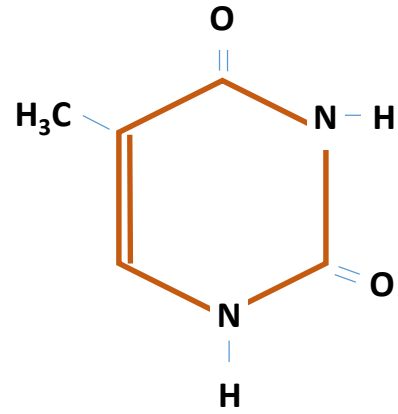


Phosphoric acid

DNA nucleobases



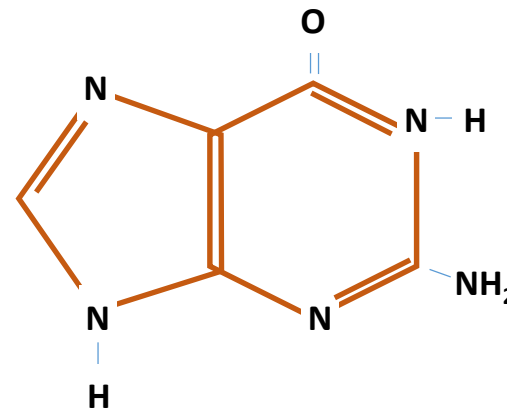
Cytosine



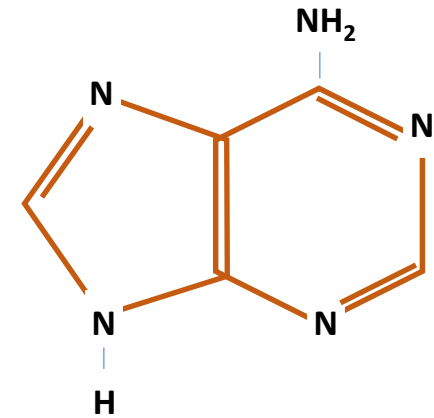
Thymine

PYRIMIDINES

PURINES

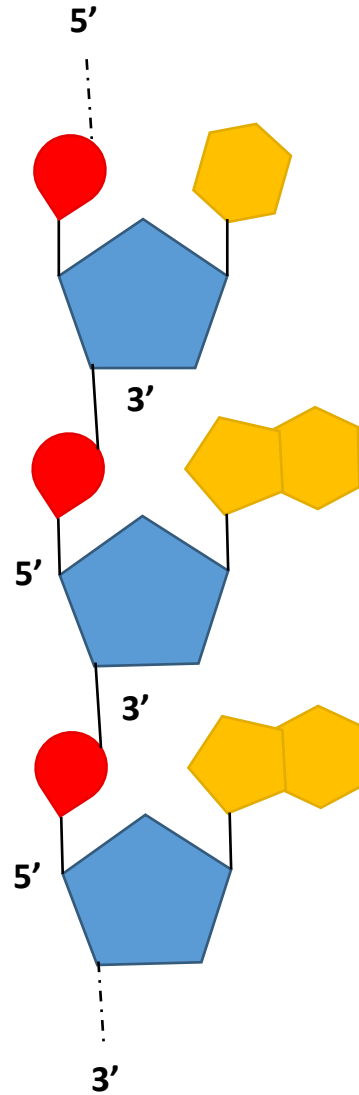


Guanine



Adenine

DNA primary structure: chain of nucleotides with phosphodiester bonds.



CHARGAFF RULES: to explain the proportion of different nucleobases in double-helix DNA.

1.- **A=T:** the number of Adenines equals the number of Thymines.

2.- **C=G:** the number of Cytosines equals the number of Guanines.

3.- **Purines=pyrimidines:** *purines* (Adenine + Guanine) = *pyrimidines* (Thymine + Cytosine).

ACGTTTTCCGACGATCTAAA
| | | | | | | | | | | | | | | | | |
TGCAAAGGCTGCTAGATTT

TTCCGTTTTCCGACTTCGCT
| | | | | | | | | | | | | | | | | |
AAGGCAAAGGCTGAAGCGA

ACGTTTCCGACGATCTAAA
| | | | | | | | | | | | | | | |
TGC AAAAGGCTGCTAGATTT

$$A/T = 12/12$$

$$C/G = 8/8$$



$$(A+G) = (T+C) = 20$$

TTCCGTTTCCGACTTCGCT
| | | | | | | | | | | | | | | |
AAGGC AAAAGGCTGAAGCGA

$$A/T = 10/10$$

$$C/G = 10/10$$



$$(A+G) = (T+C) = 20$$

CHARGAFF RULES: to explain the proportion of different nucleobases in double-helix DNA.

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There is a purine for each pyrimidin, and they appear coupled

CHARGAFF RULES: to explain the proportion of different nucleobases in double-helix DNA.

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4.- **(A+T)/(C+G):** Adenine & Thymine and Cytosine & Guanine proportion is variable and depends on the species considered.

ACGTTTCCGACGATCTAAA
| | | | | | | | | | | | | | | |
TGC AAAAGGCTGCTAGATTT

$$A/T = 12/12$$

$$C/G = 8/8$$



$$(A+T)/(C+G) = 24/16$$

TTCCGTTTCCGACTTCGCT
| | | | | | | | | | | | | | | |
AAGGC AAAAGGCTGAAGCGA

$$A/T = 10/10$$

$$C/G = 10/10$$



$$(A+T)/(C+G) = 20/20$$

CHARGAFF RULES: to explain the proportion of different nucleobases in double-helix DNA.

1.- **A=T:** the number of Adenines equals the number of Thymines.

2.- **C=G:** the number of Cytosines equals the number of Guanines.

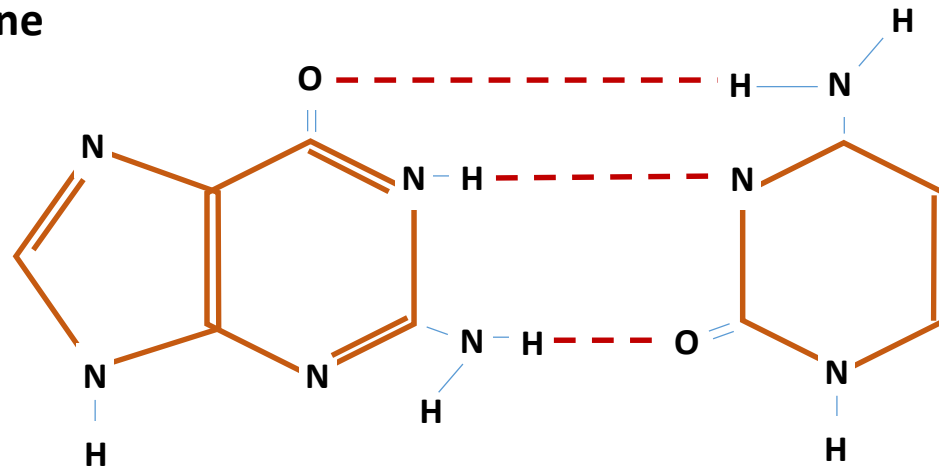
3.- **Purines=pyrimidines:** *purines* (Adenine + Guanine) = *pyrimidines* (Thymine + Cytosine).

There is a purine for each pyrimidin, and they appear coupled

4.- **(A+T)/(C+G):** Adenine & Thymine and Cytosine & Guanine proportion is variable and depends on the species considered.

DNA molecules differ in nucleobases composition

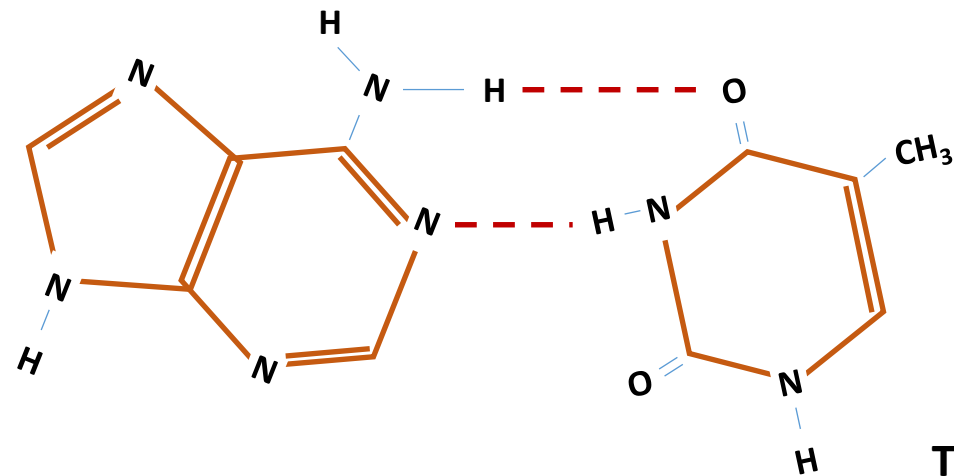
Guanine



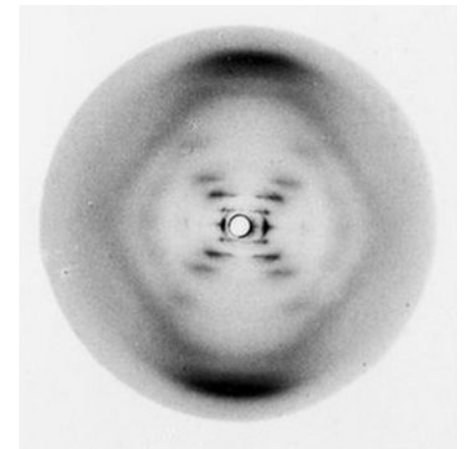
Complementarity

Cytosine

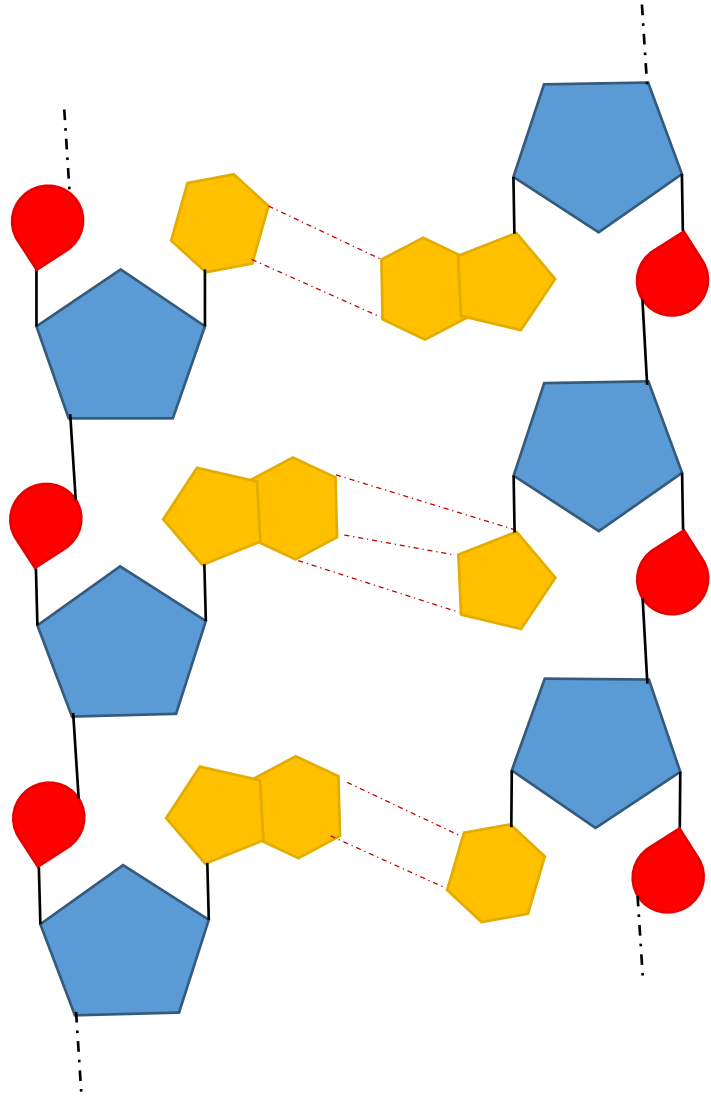
Adenine

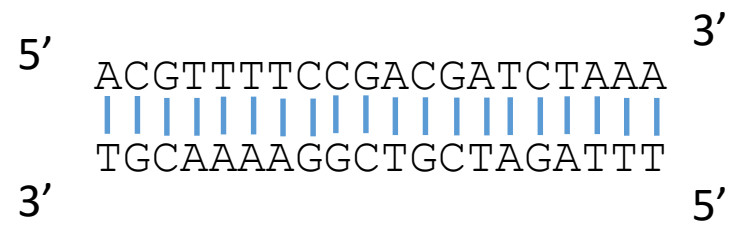


Thymine

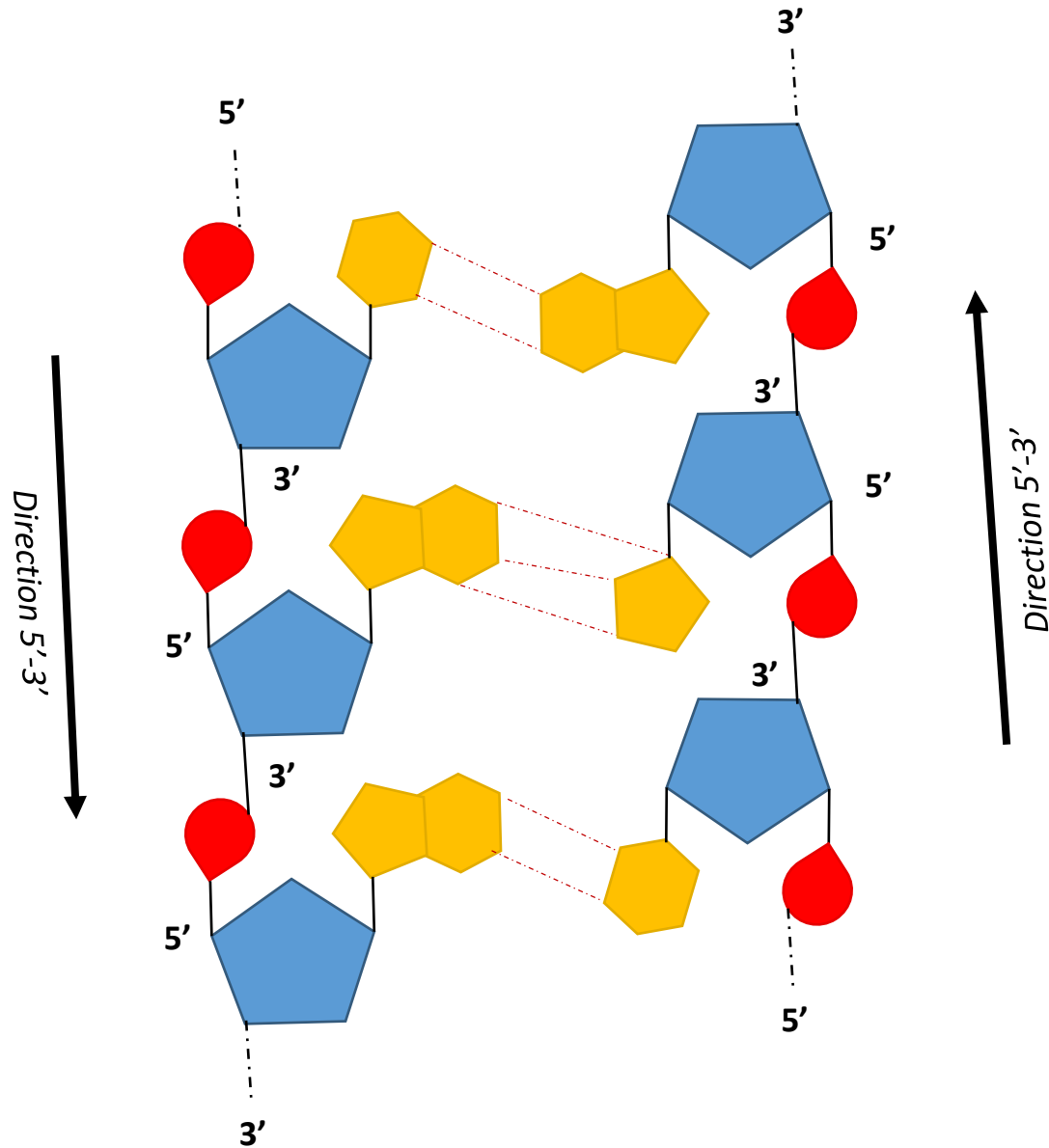


DNA secondary structure: chains are antiparallel and complementary.



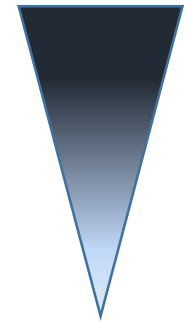


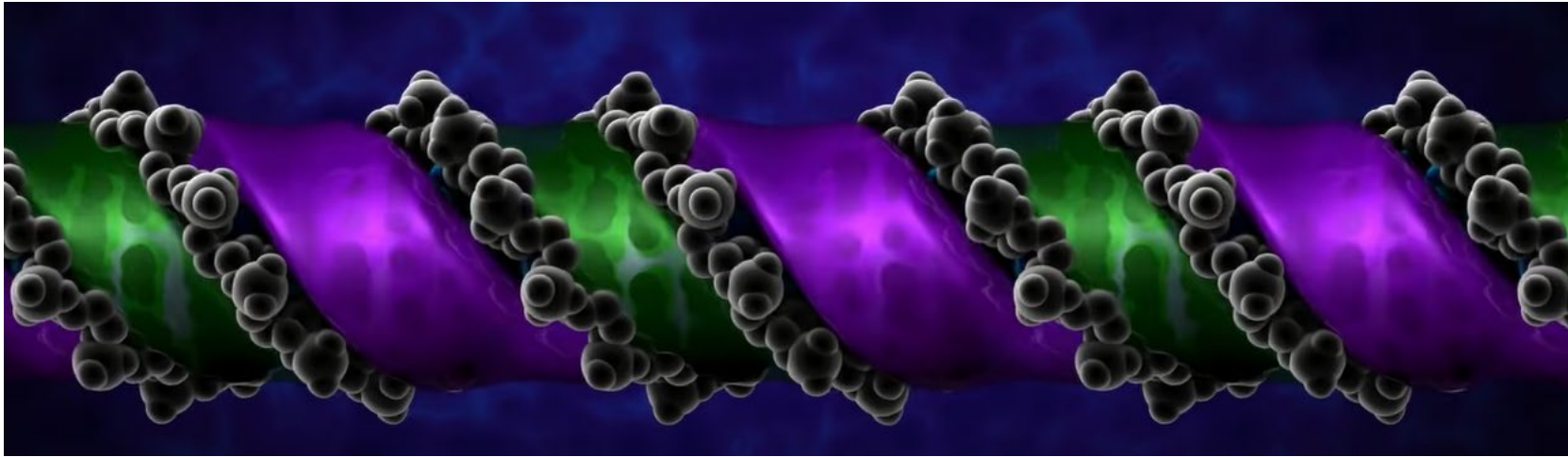
DNA secondary structure: chains are antiparallel and complementary.



TYPES OF BONDS:

- Phosphodiester (1^a)
- H bond(2^a)
- Pi-stacking (2^a)

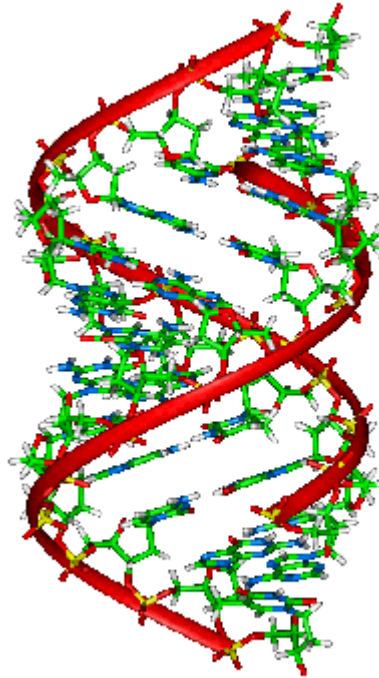




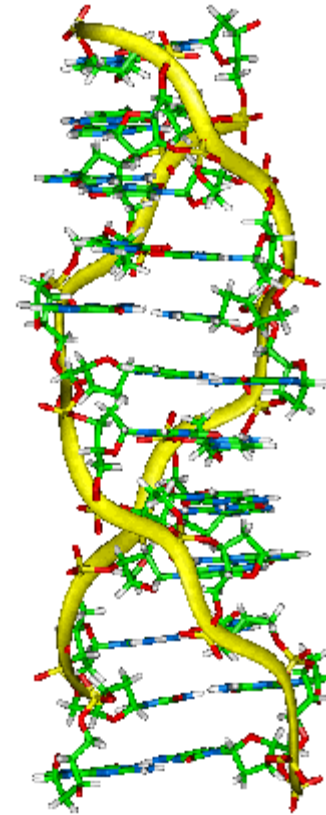
DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.



DNA-B

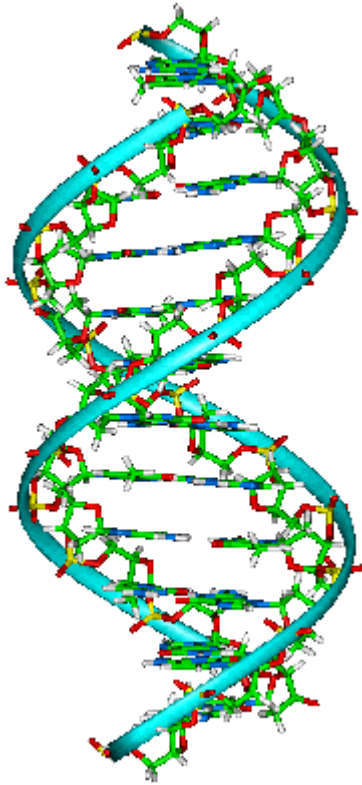


DNA-A



DNA-Z

DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.

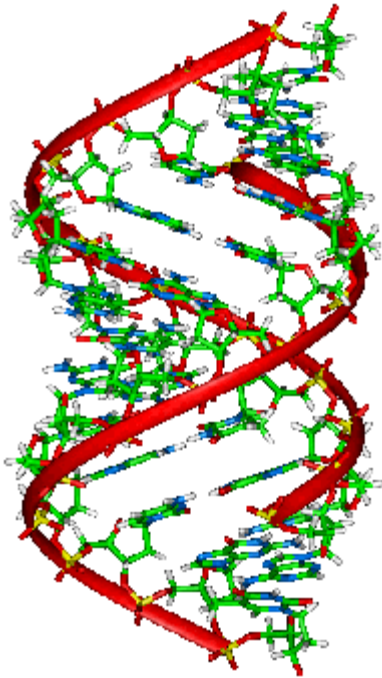


DNA-B

Particulars	B DNA
Helix	Right handed
Base pairs per turn	~10.5
Helical Diameter (nm)	2.0
Helical length (nm)	3.4
Shape	Intermediate
Major Grove	Narrow, deep
Minor Grove	Broad, shallow

B-DNA
92% H₂O

DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.



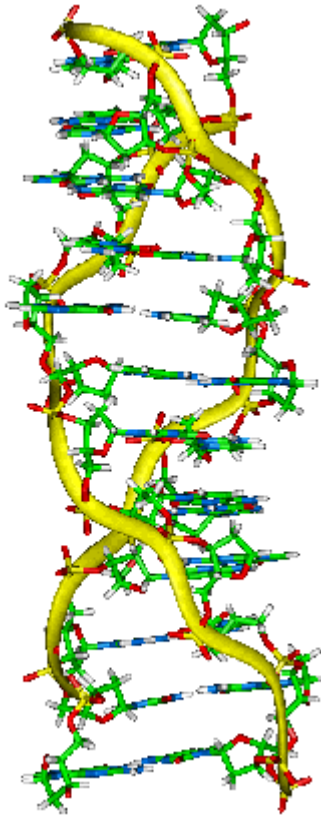
DNA-A

Particulars	A DNA
Helix	Right handed
Base pairs per turn	~11
Helical Diameter (nm)	2.6
Helical length (nm)	2.6
Shape	Broadest
Major Grove	Wide, deep
Minor Grove	Narrow, shallow

A-DNA

75% H₂O

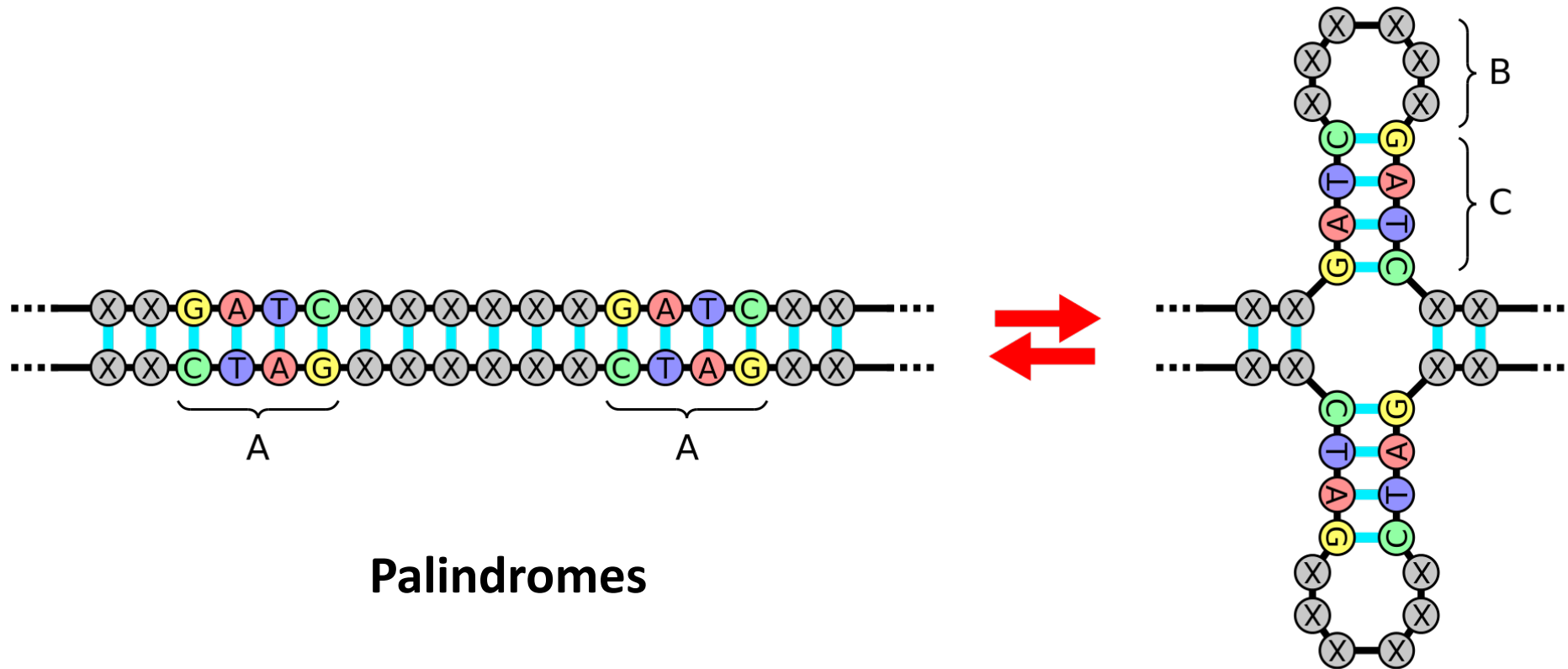
DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.



DNA-Z

Particulars	Z DNA
Helix	Left handed
Base pairs per turn	~12
Helical Diameter (nm)	1.8
Helical length (nm)	3.7
Shape	Narrowest
Major Grove	Flat
Minor Grove	Narrow, deep

DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.

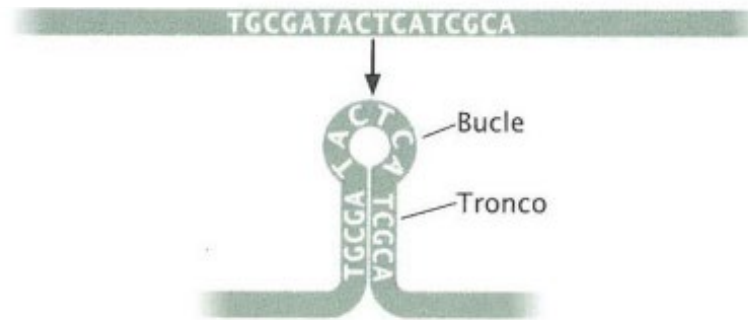


Palindromes

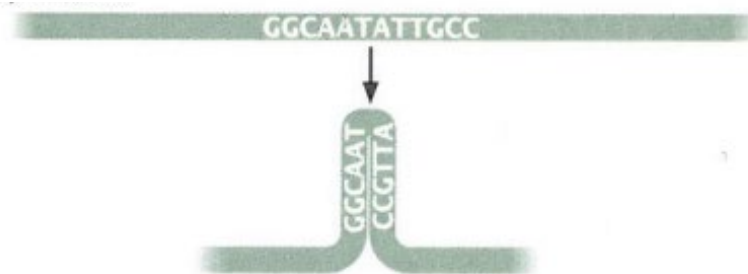
DNA secondary structure: chains are antiparallel and complementary.
Stable 3D structure.

Palindromes & Complementary Regions

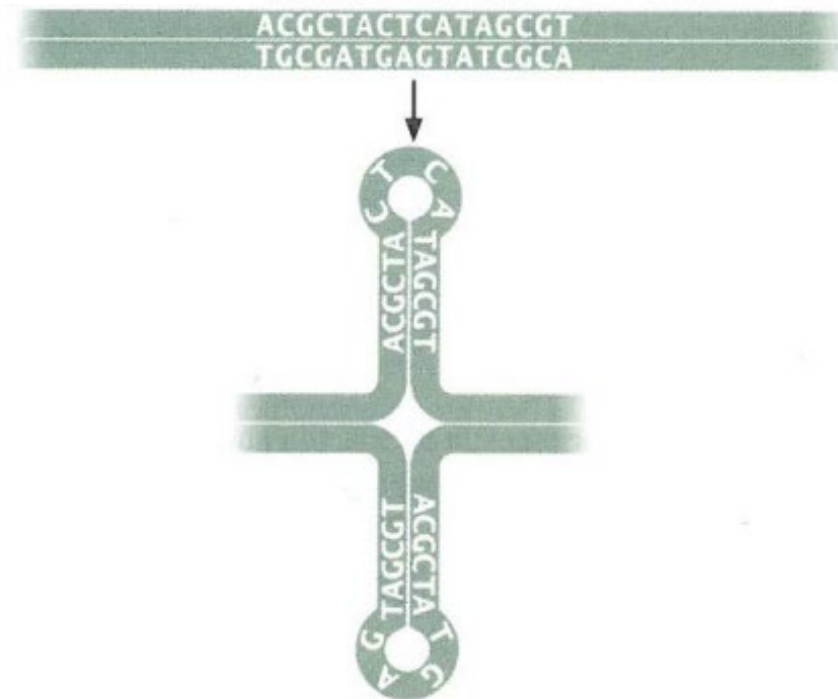
Loops



Forks

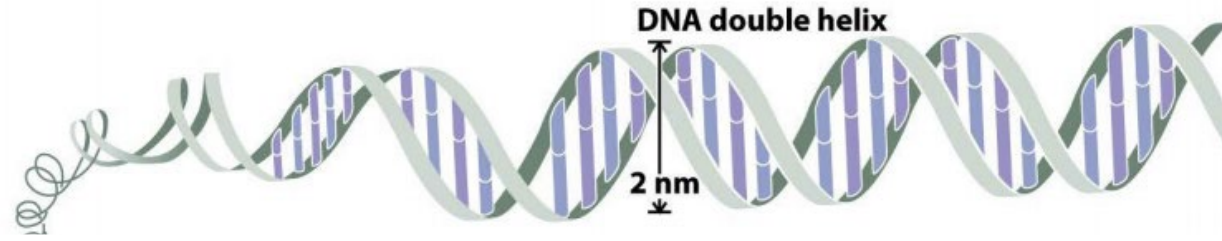


Cross

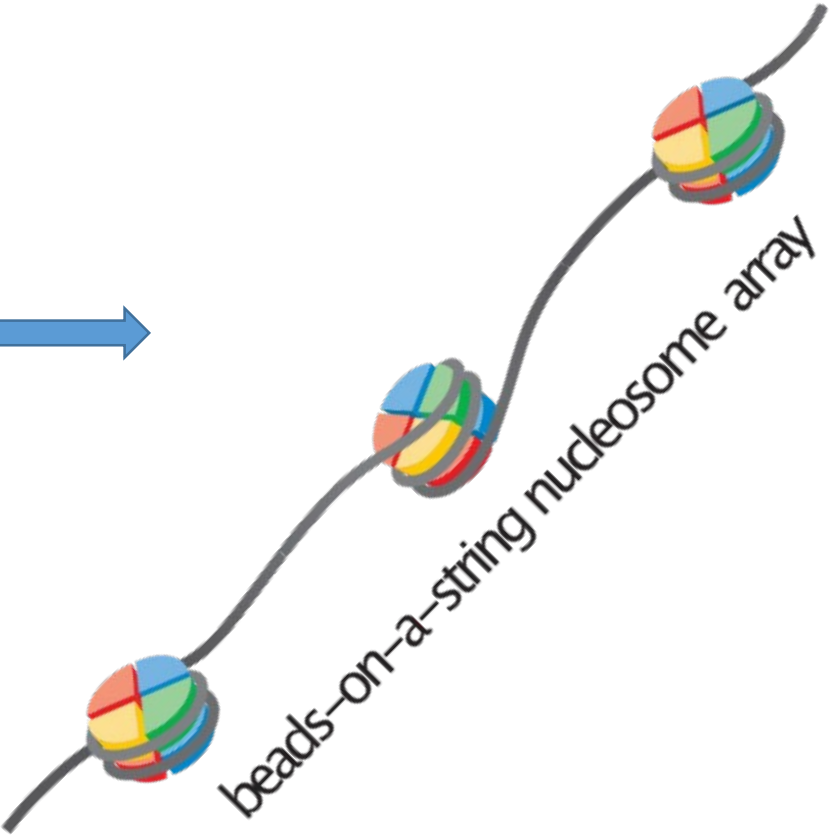
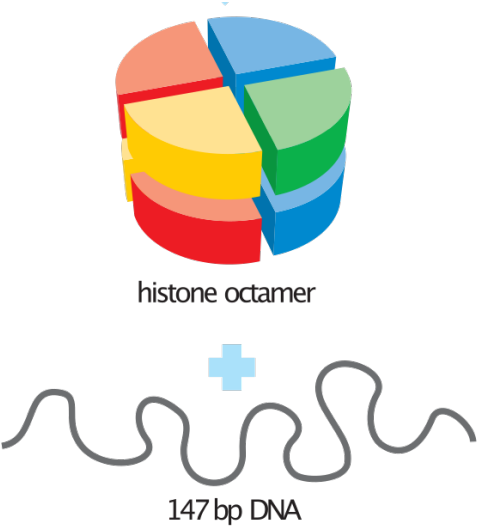
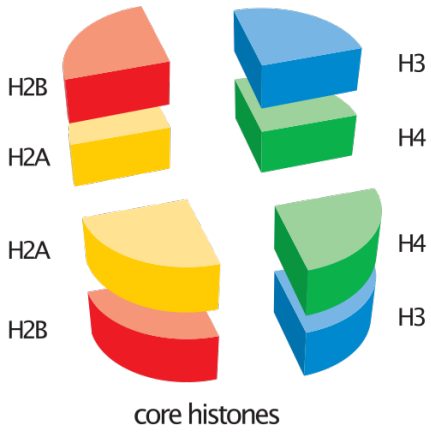


(c)

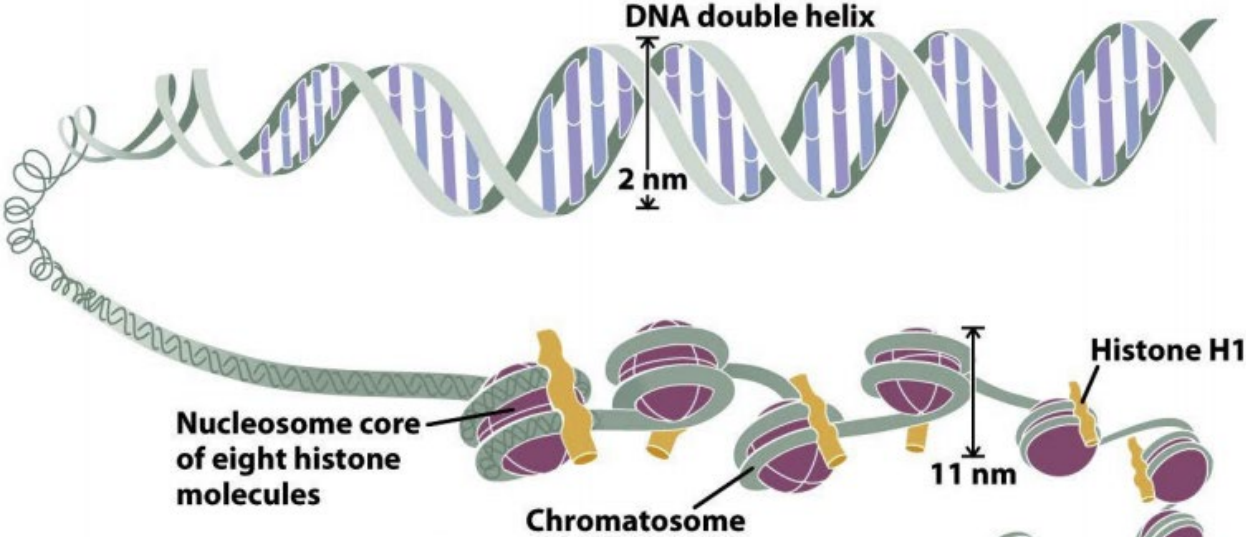
DNA tertiary structure: DNA packaging and protein binding.

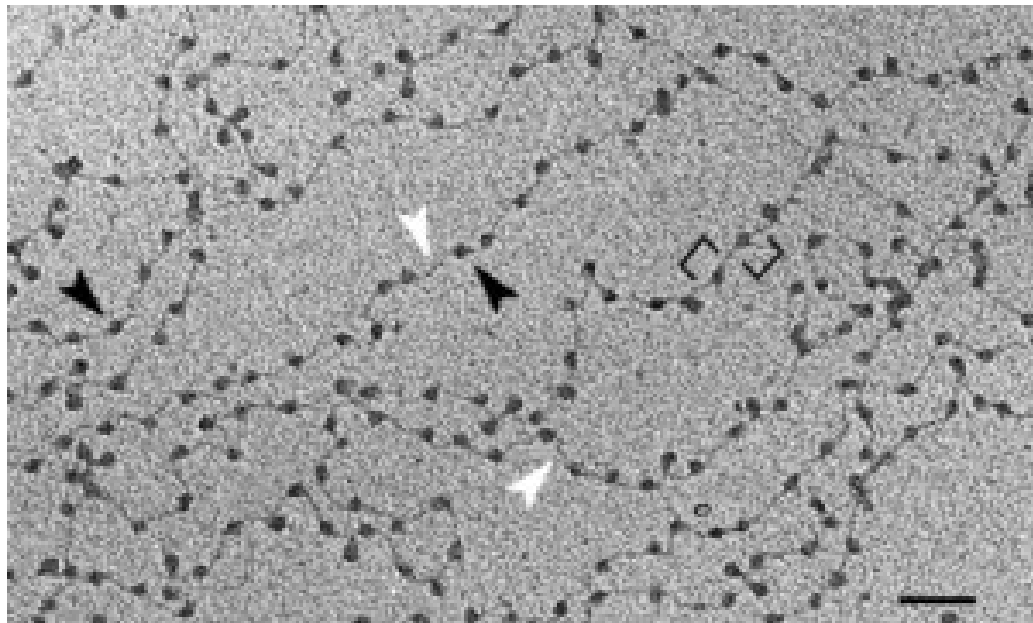
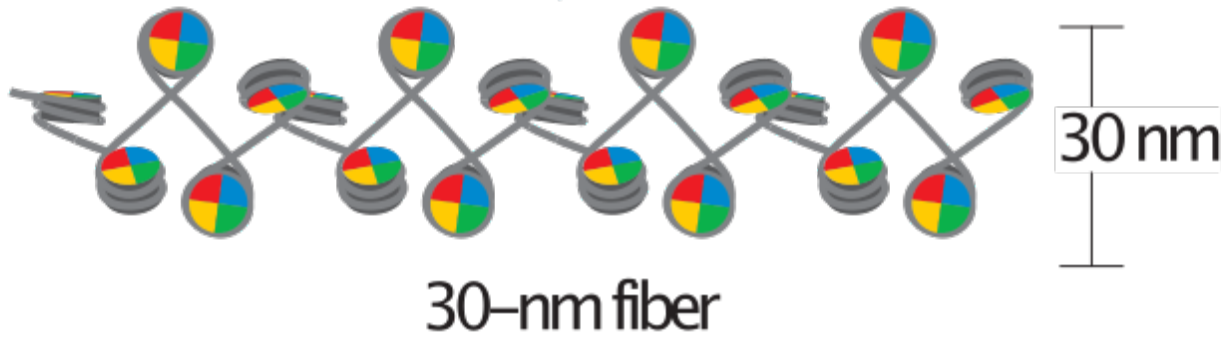


Histone Octamer or Nucleosome

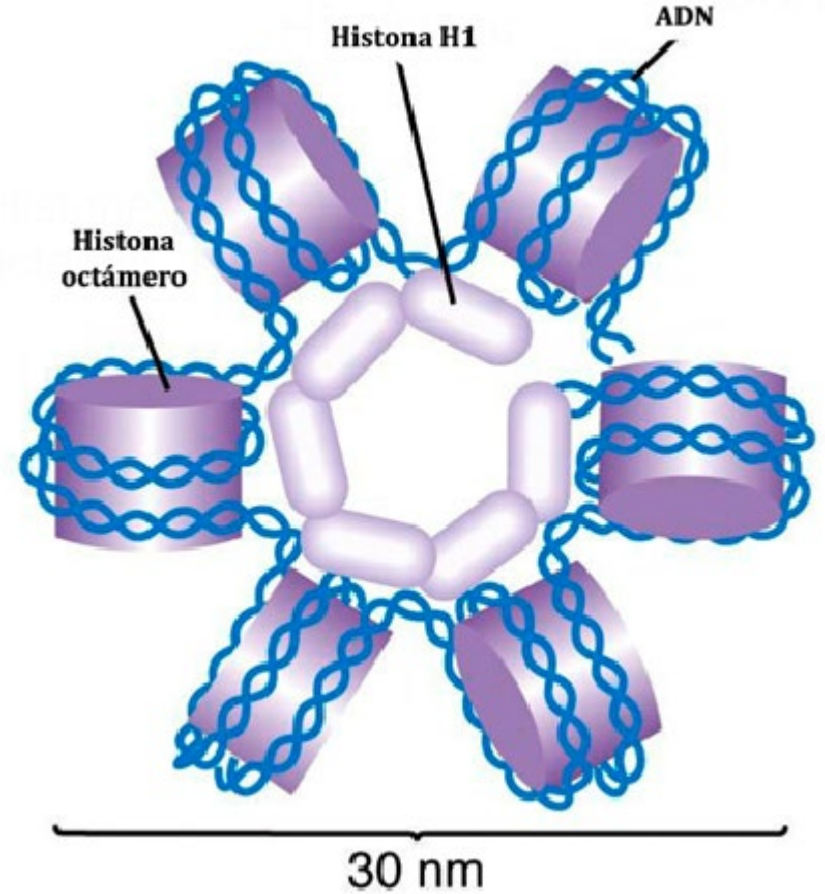


DNA tertiary structure: DNA packaging and protein binding.

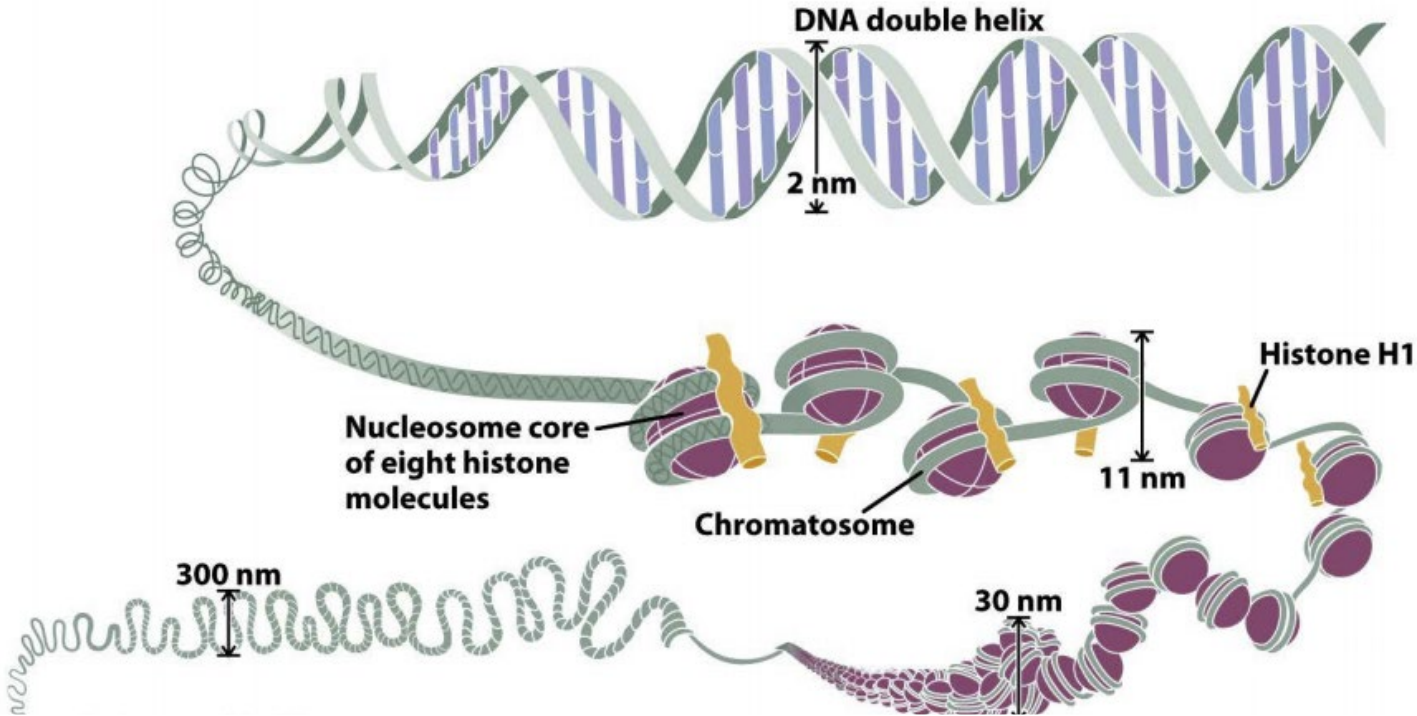




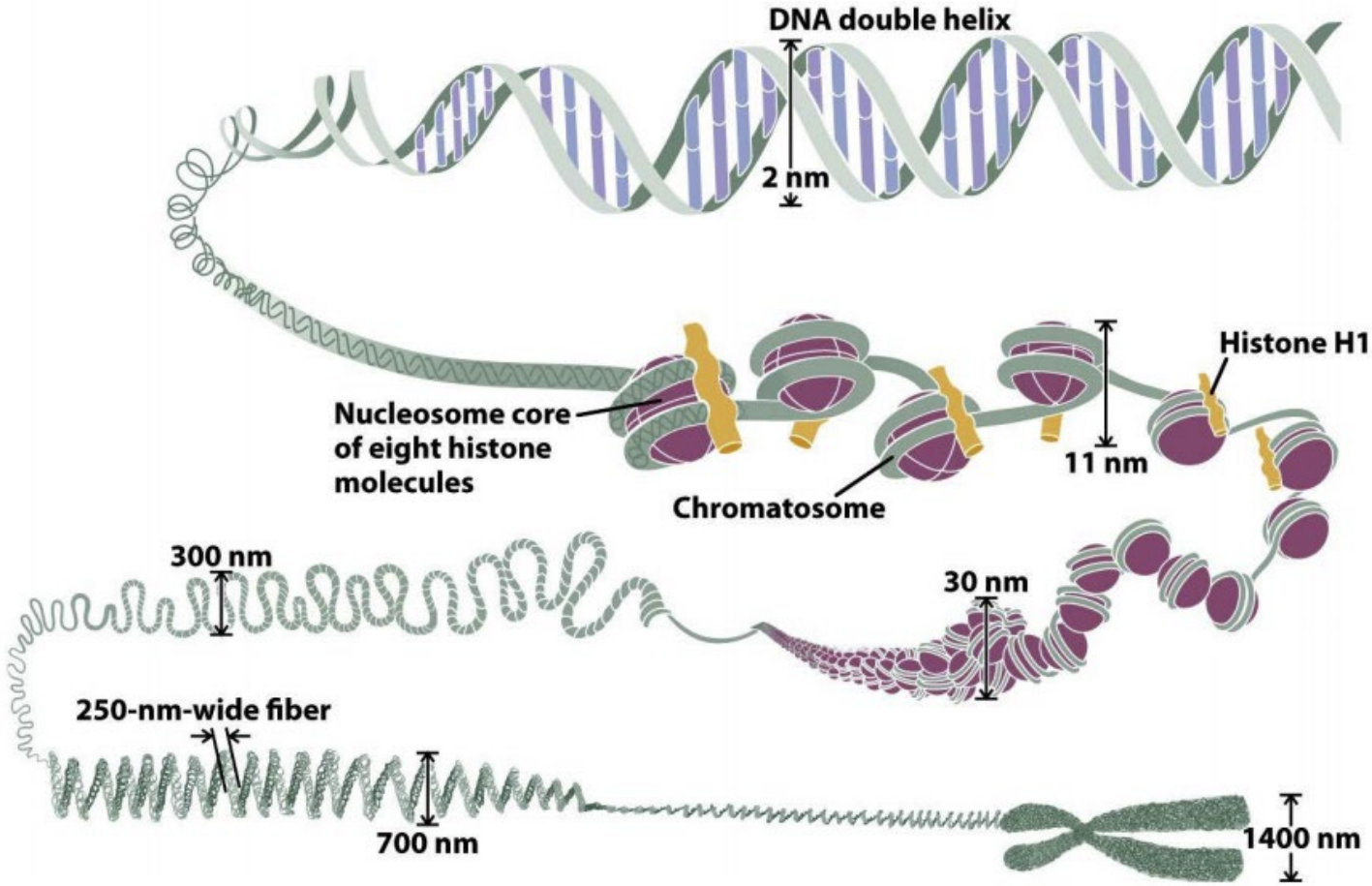
Beads on "string" structure



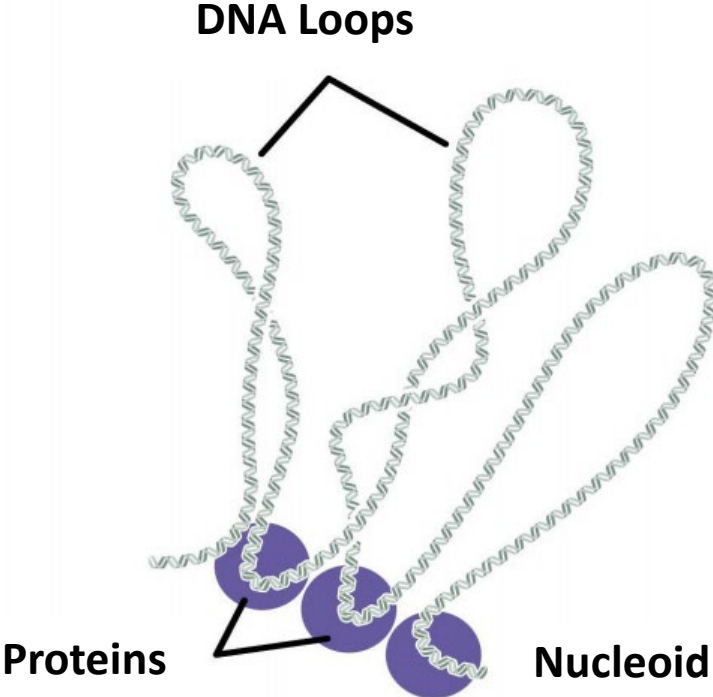
DNA tertiary structure: DNA packaging and protein binding.



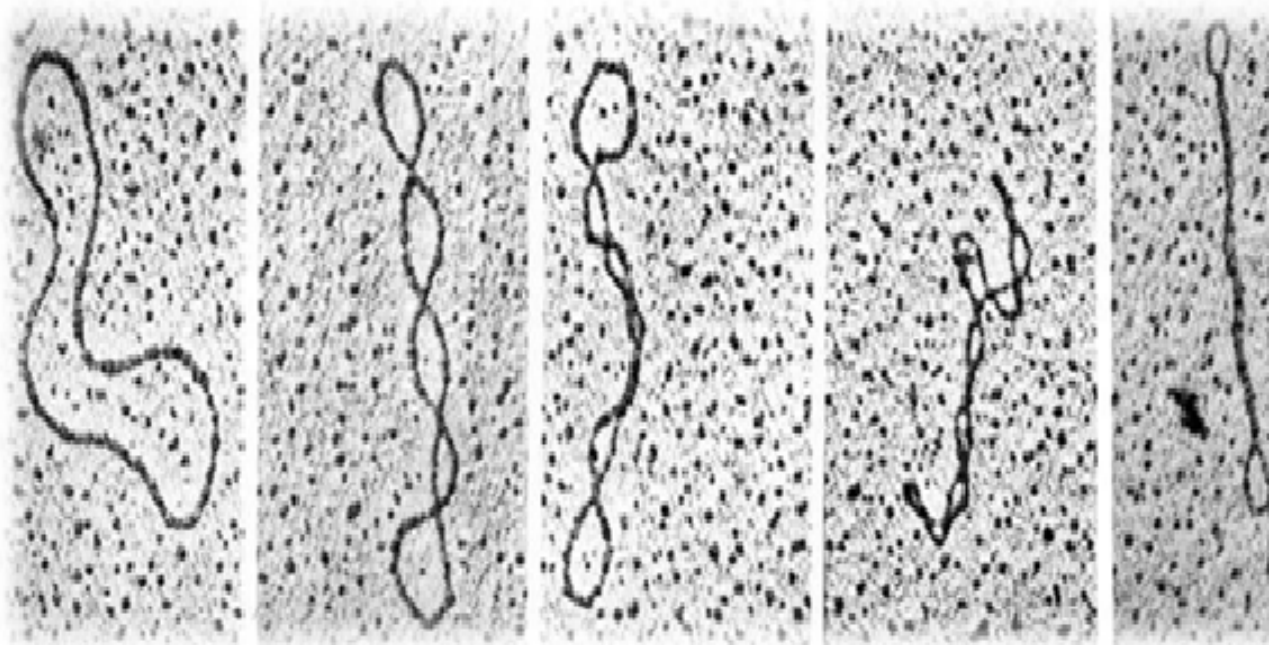
DNA tertiary structure: DNA packaging and protein binding.



DNA tertiary structure: DNA packaging and protein binding.



DNA SUPERCOILING IN BACTERIAL CHROMOSOMES:



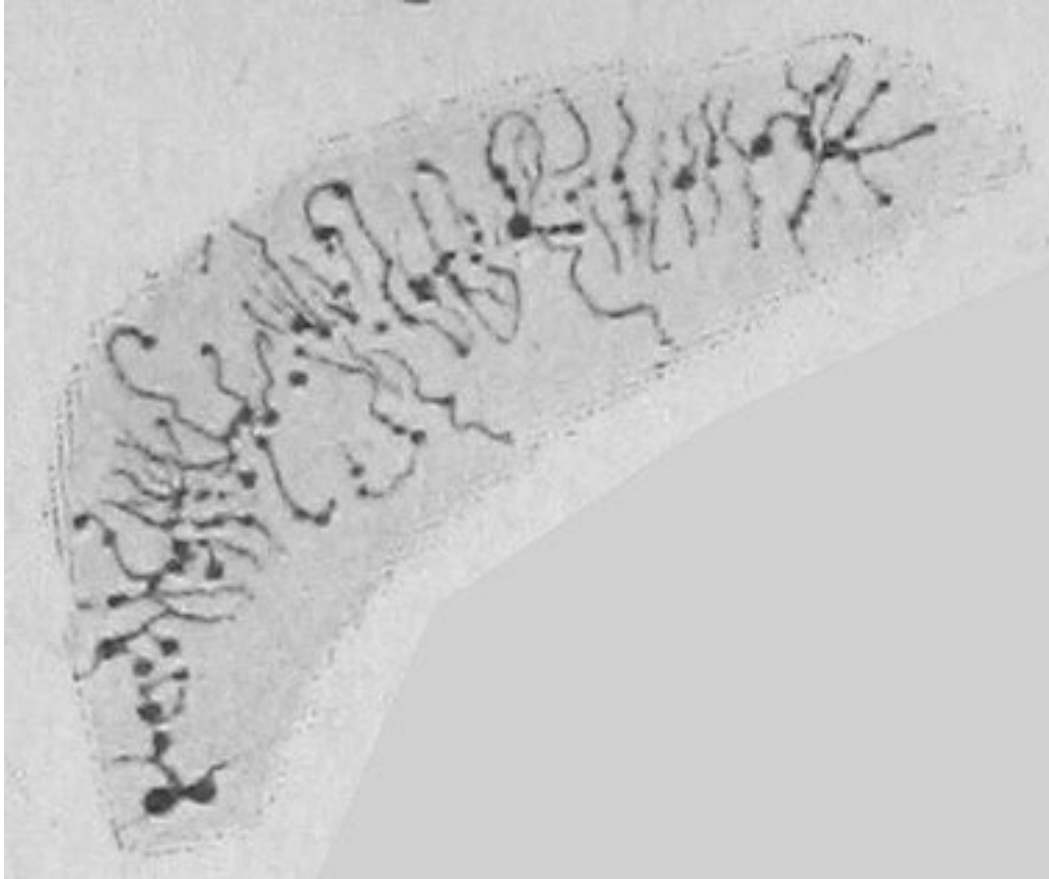
DNA tertiary structure: DNA packaging and protein binding.



POLYTENE CHROMOSOMES O CROMOSOMAS GIGANTES:

- In salivary glands, digestive tract and rectum of *Drosophila* larvae.
- Homologous chromosomes are paired (**although they belong to somatic cells**).
- Multiple DNA strands paired.
- Intensely stained regions and decondensed regions (*puffs*), these later indicate high rate of transcription.

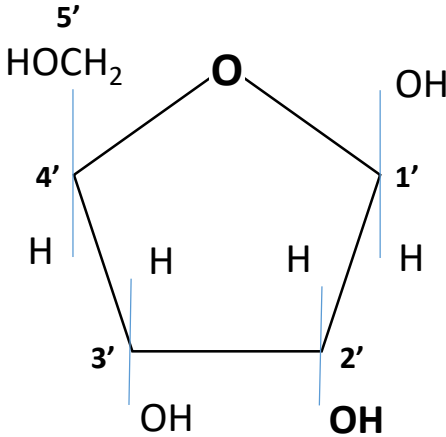
DNA tertiary structure: DNA packaging and protein binding.



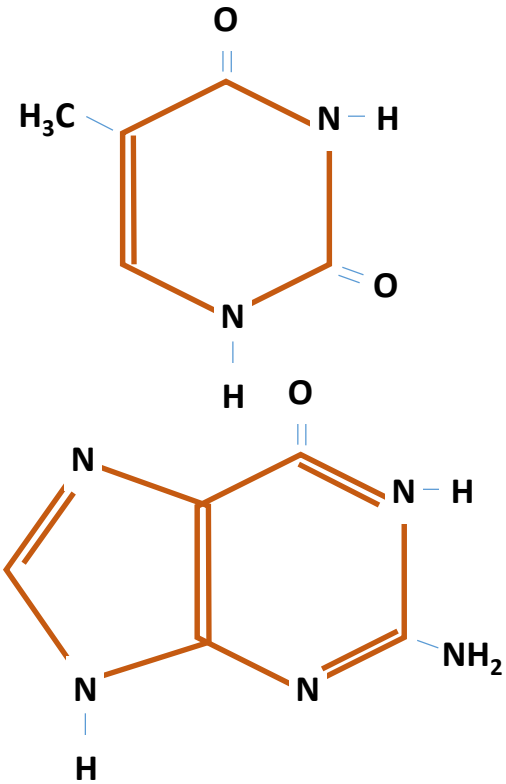
LAMPBRUSH CHROMOSOMES:

- Oocytes of sharks, salamanders and spermatocytes of many vertebrates.
- Meiotic chromosomes with regions of decondensed chromatin.
- Large size, especially in diplotene.
- These *loops* are probably made of RNA (indicating high rate of gene transcription).

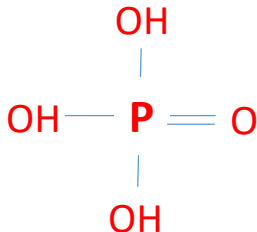
RNA composition



Pentose
(Ribose)

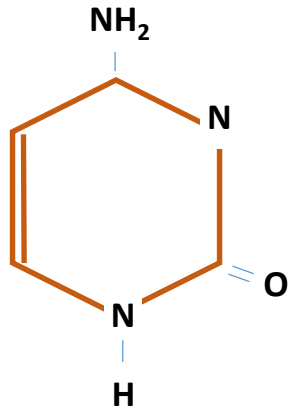


Nucleobase

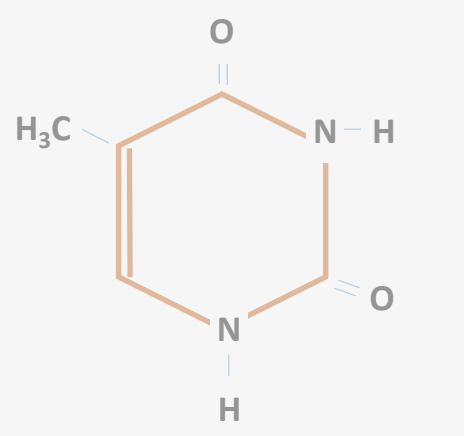


Phosphoric acid

RNA nucleobases

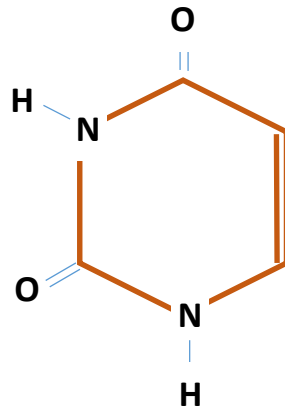


Cytosine



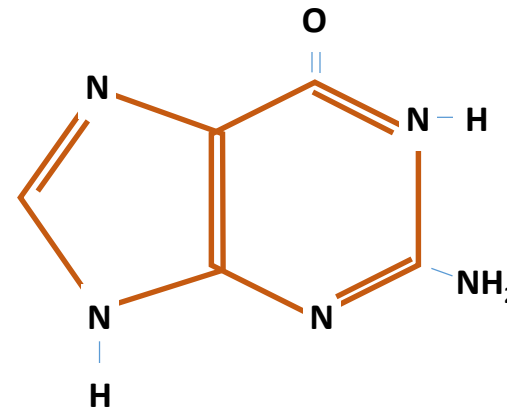
Thymine

PYRIMIDINES

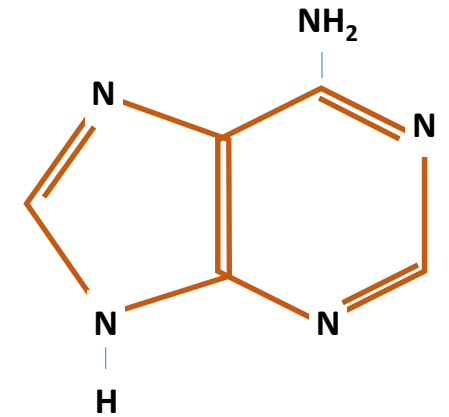


Uracil

PURINES

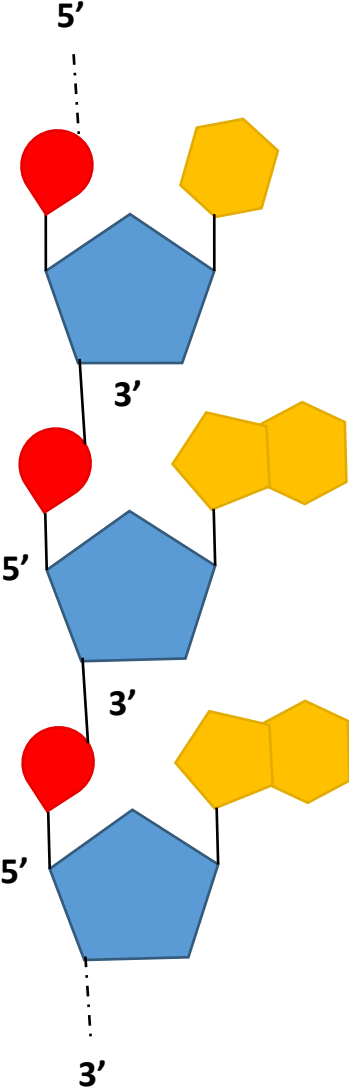


Guanine

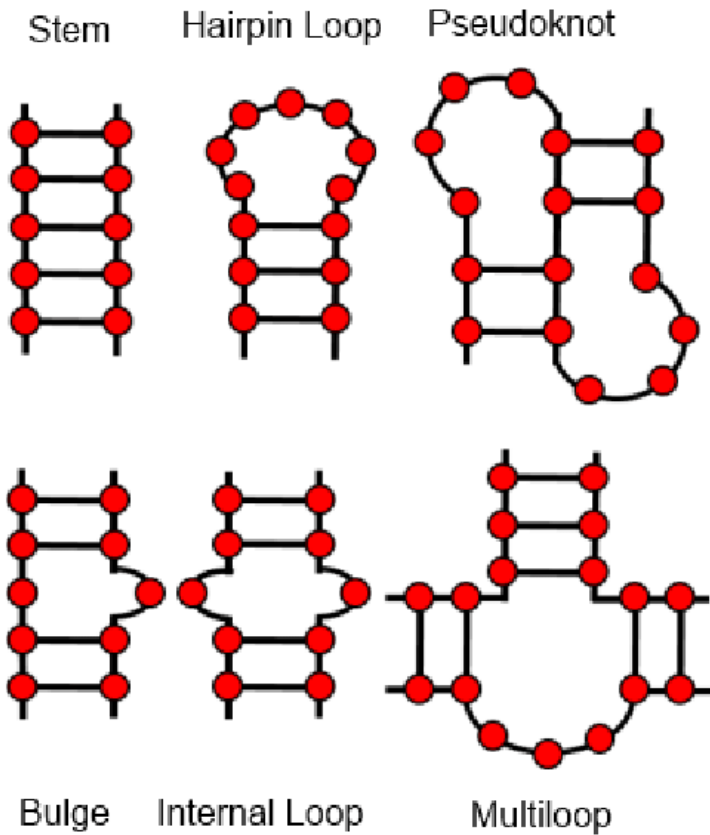


Adenine

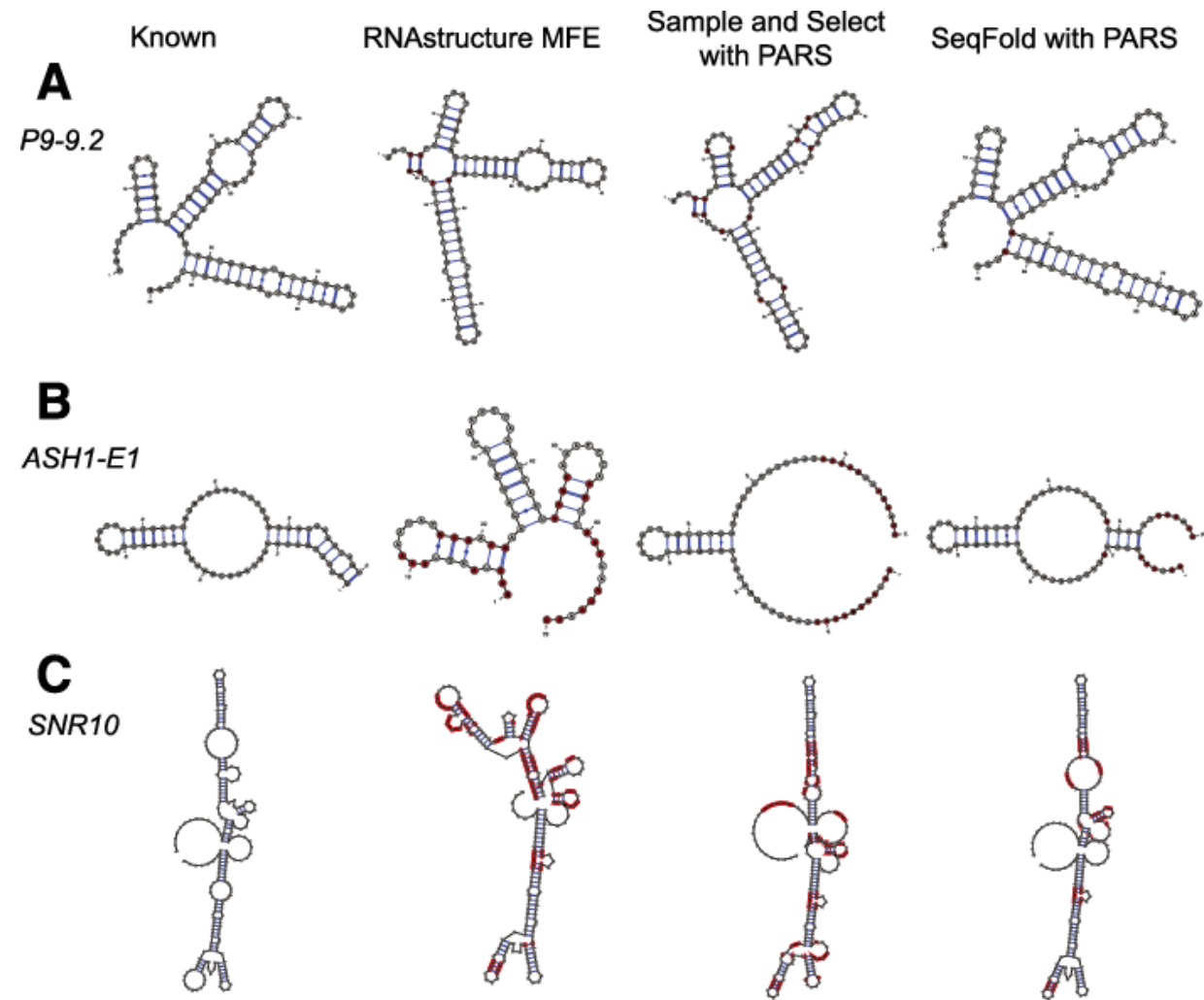
RNA primary structure: chain of nucleotides with phosphodiester bonds.



RNA secondary structure

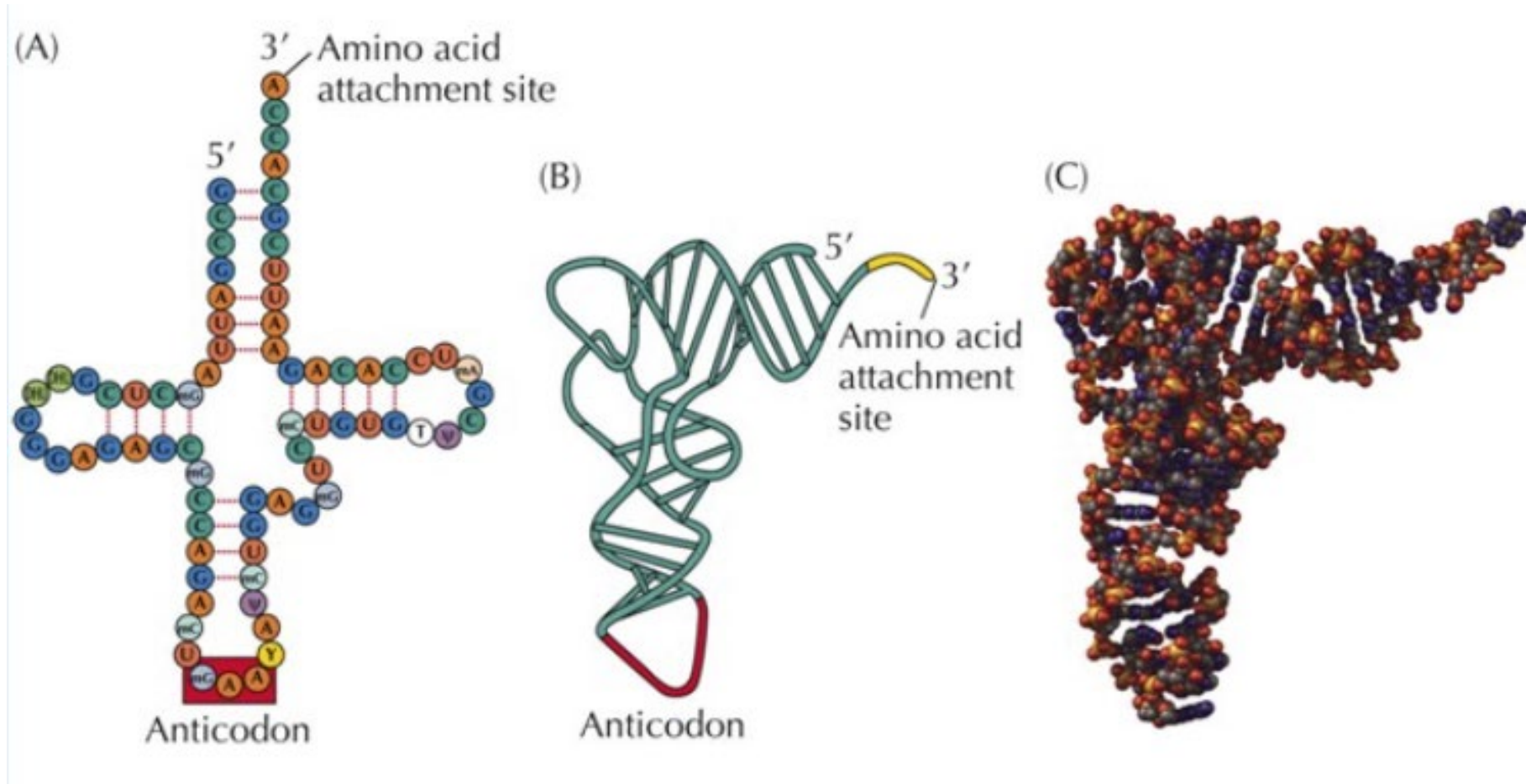


Source: Manish Kumar Gupta, ResearchGate

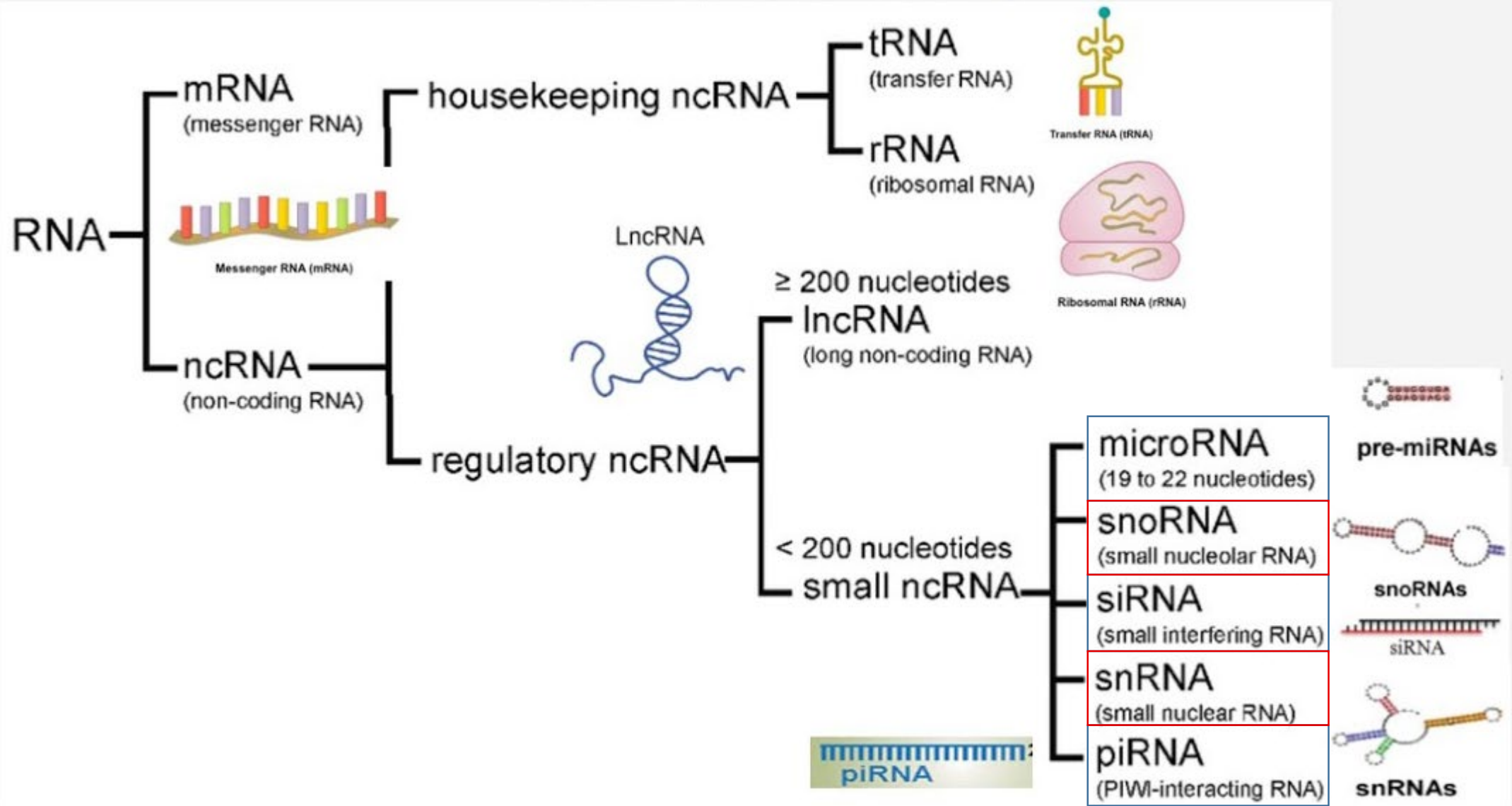


Source: Zhengqing Ouyang, ResearchGate

RNA tertiary structure: tRNA



Main types of RNA



Main differences DNA/RNA

Bases	A, G, C, T	A, G, C, U
Strand	Double stranded	Single stranded
Structure	Antiparallel helix	Hairpin and loops
Sugar	Deoxyribose	Ribose
Location	Nuclear or mitochondrial	Nuclear or cytoplasmic
Lifetime	Long	Short
Process	Transcription	Translation
Types	Nuclear DNA, mtDNA	mRNA, tRNA, rRNA, miRNA, siRNA, ribozyme

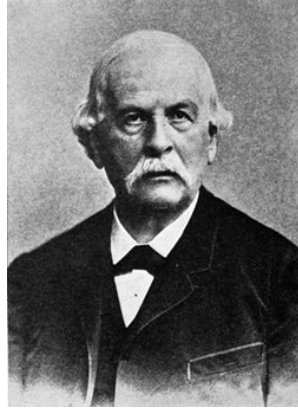
Features of hereditary material

- **Stable way to store information.** Instructions for all traits and functions.
- **Auto-replicate with fidelity.**
- **Express the information.** Central Dogma of Biology DNA->RNA->Protein

Pioneers in DNA structure and composition



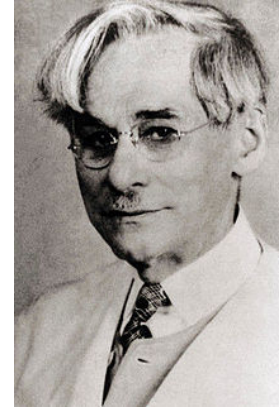
Friedrich Miescher,
1869



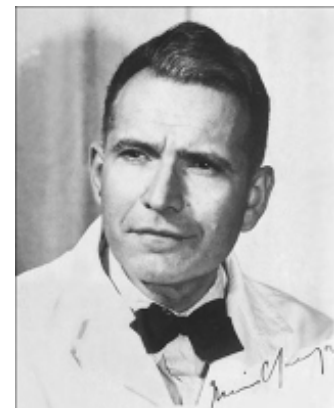
Richard Altmann,
1889



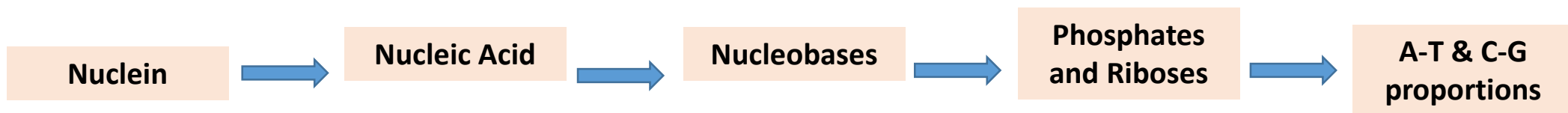
Albrecht Kossel,
1910



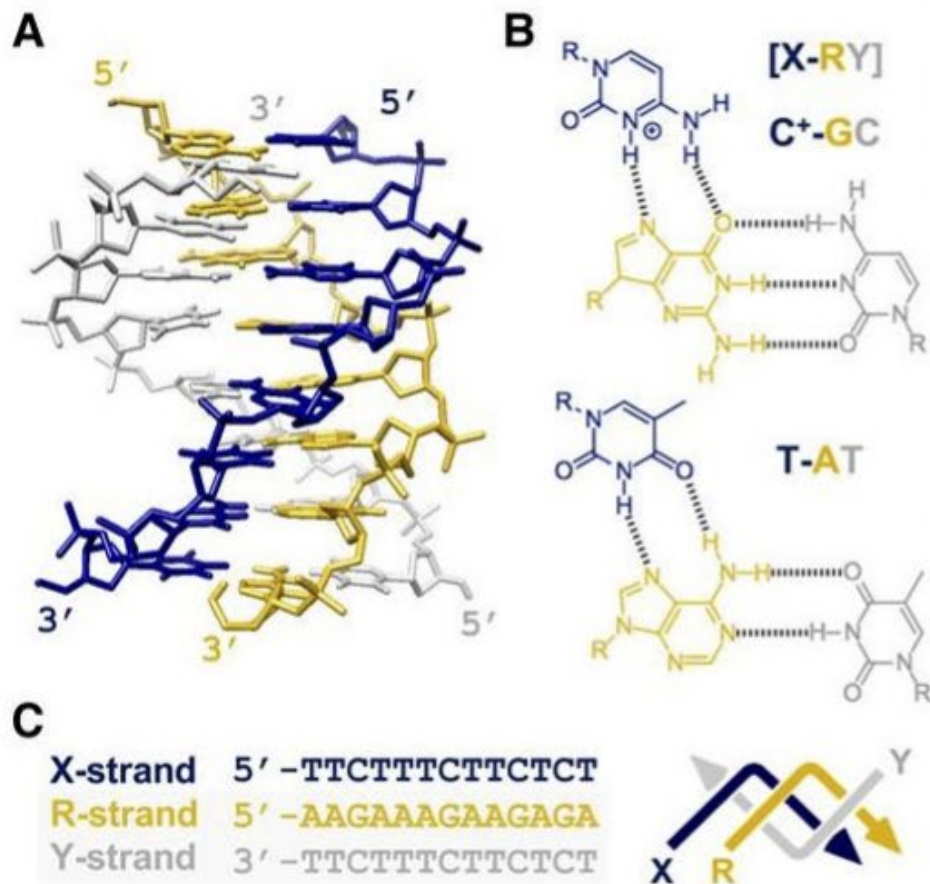
Phoebus Levene,
1920



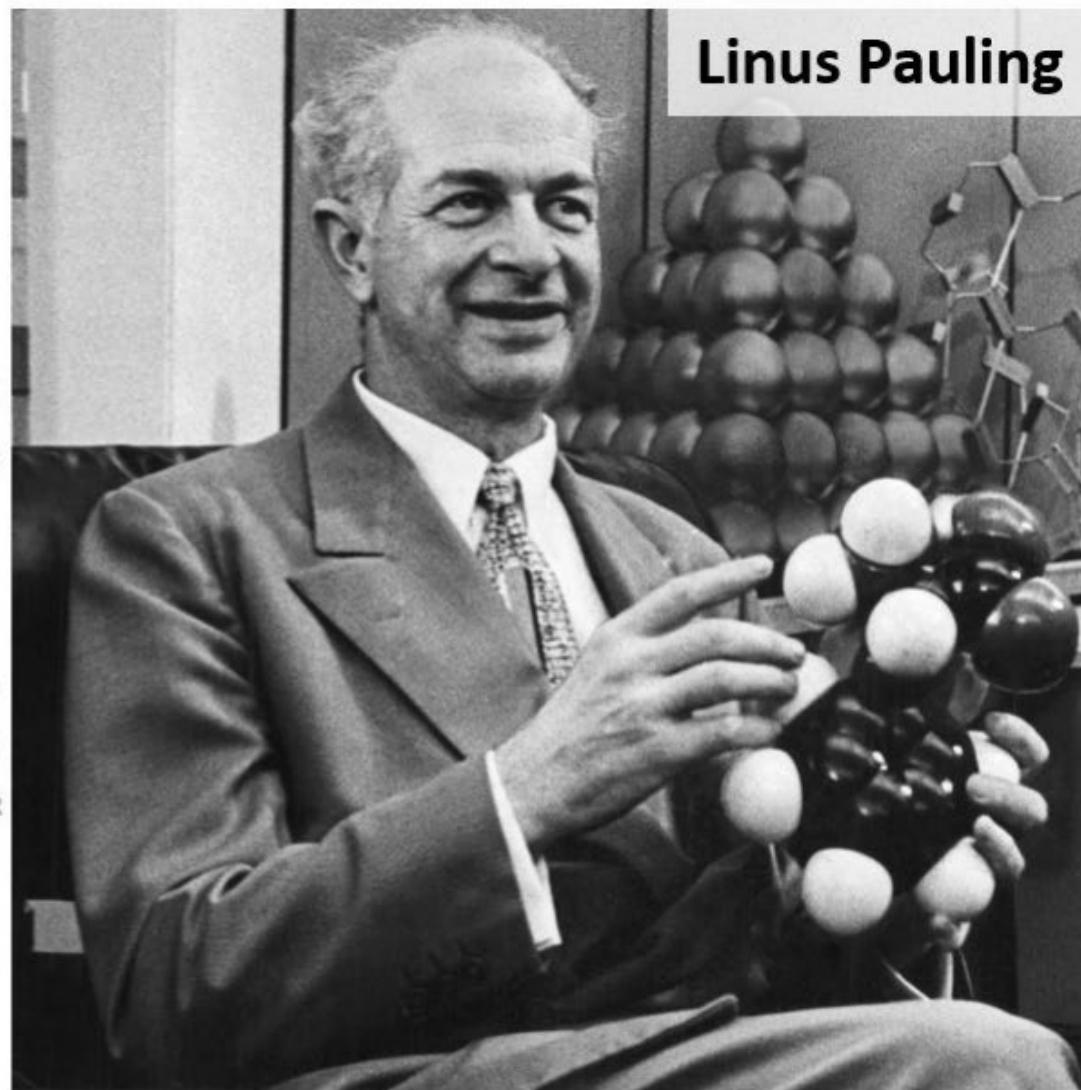
Erwin Chargaff,
1940



Triple Helix



<https://doi.org/10.1093/nar/gkx1230>



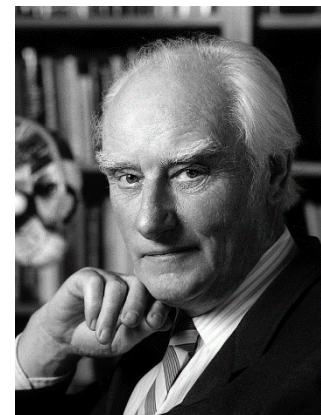
Pioneers in DNA structure and composition



Rosalind Franklin, Maurice Wilkins,
1953



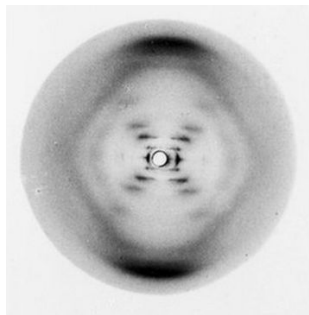
1953



Francis Crick,
1953



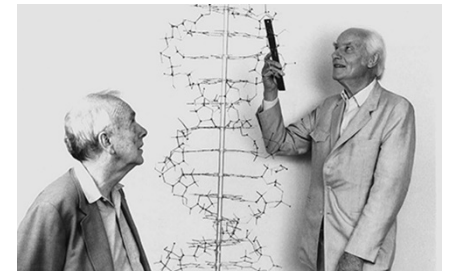
James Watson,
1953



X-ray
crystallography



Tridimensional
structure



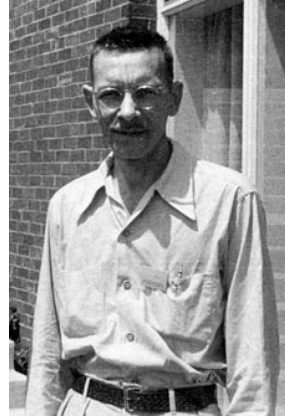
Pioneers in DNA function



Frederick Griffith,
1928



Oswald Avery,
1944



Alfred Hershey,
1952



Martha Chase,
1952

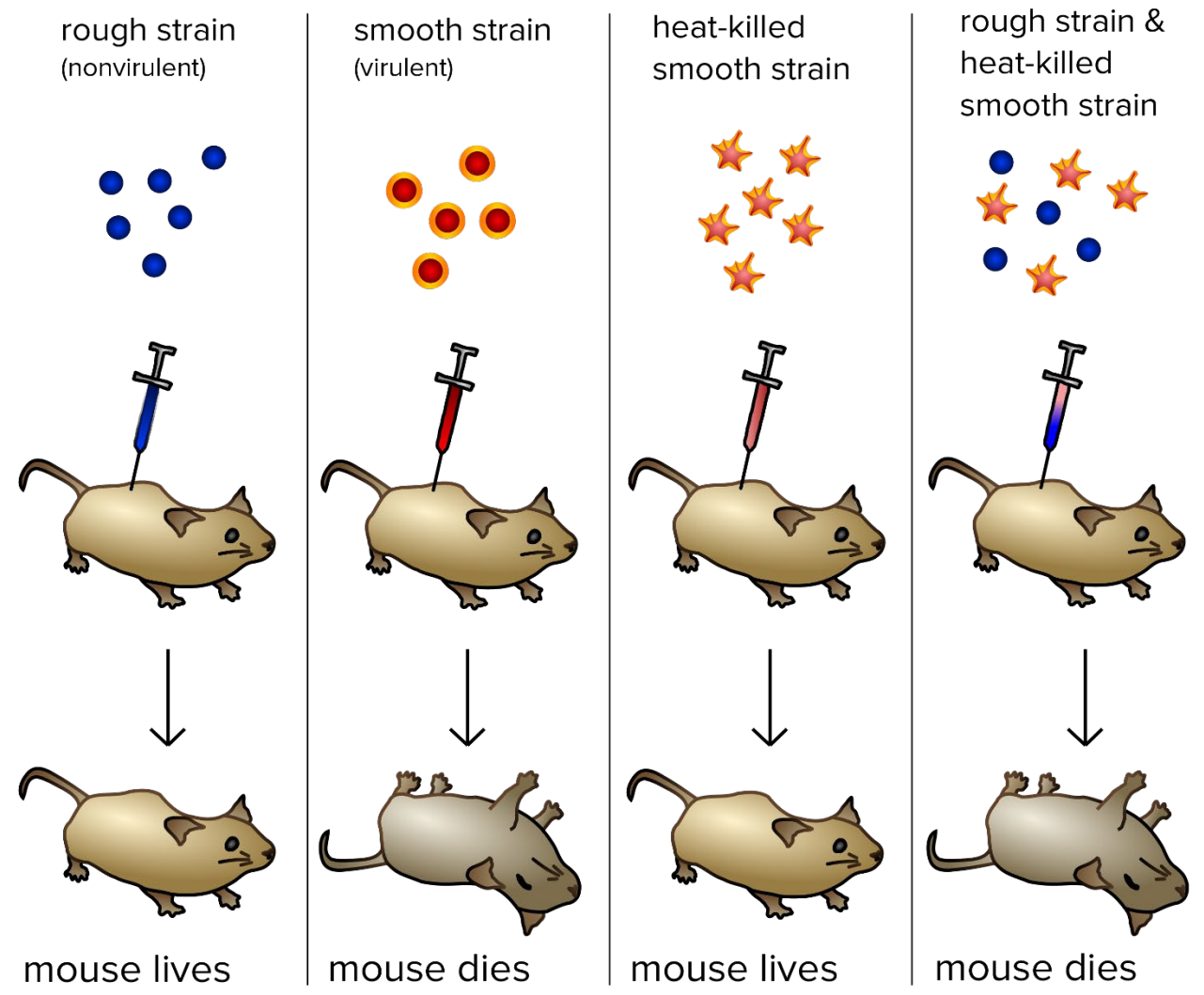
**Transformation
of bacteria**



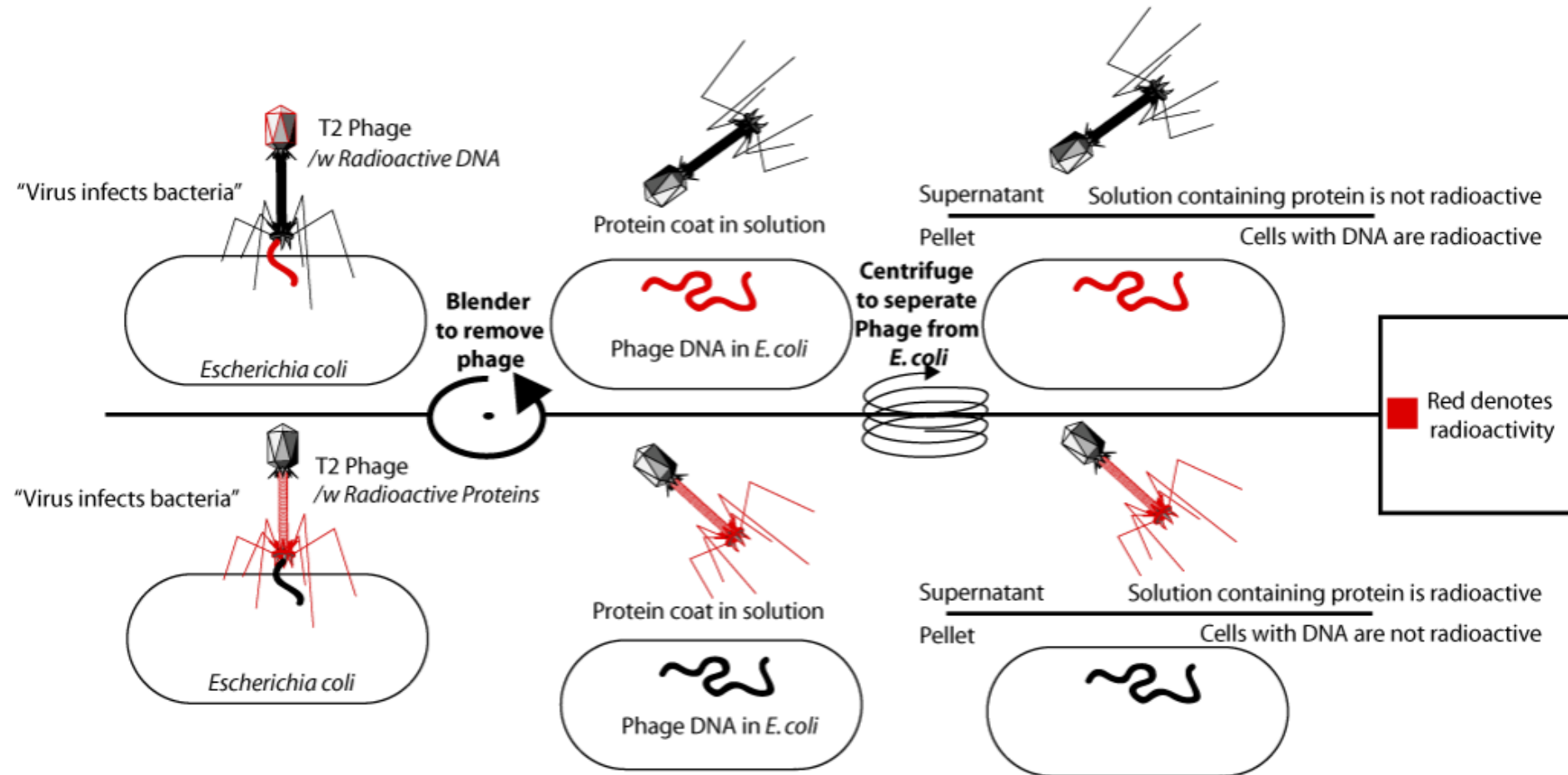
**The Transforming
Principle**



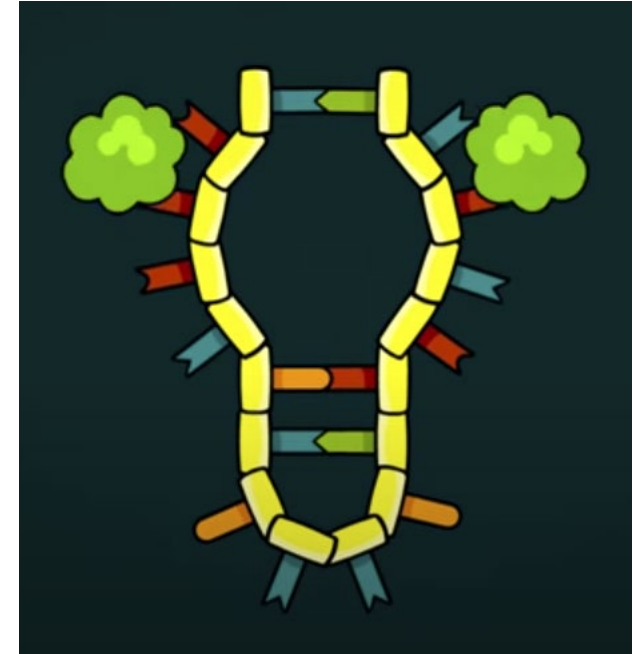
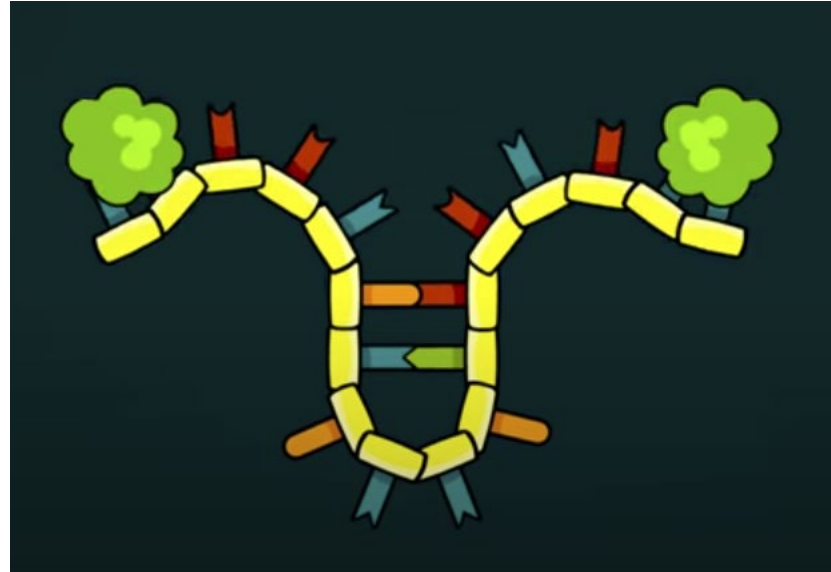
Bacteriophages



Hershey & Chase Experiment



RNA World Hypothesis



- **LUCA** (Last Universal Common Ancestor).
- RNA strands would have served as templates for **auto-replication**.
- Catalytic activity of **ribozymes** (autocatalytic and non-autocatalytic) would allow the acquisition of new “functions”.

Baltimore Classification of Virus

DNA VIRUS:

GROUP I: double strand

GROUP II: one strand

RNA VIRUS:

GROUP III: double strand

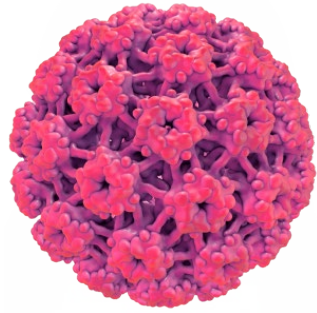
GROUP IV: one strand (+)

GROUP V: one strand (-)

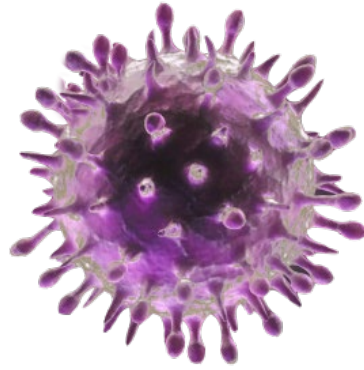
DNA VIRUS OR RNA REVERSE TRANSCRIPTASE:

GROUP VI: RNA one strand

GROUP VII: DNA double strand

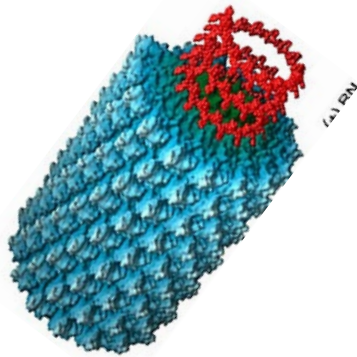


Human papilloma virus

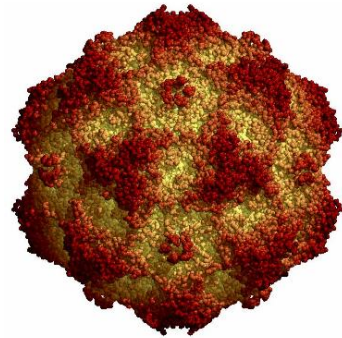


Chickenpox virus
Herpes virus

DNA VIRUS:
GROUP I: double strand

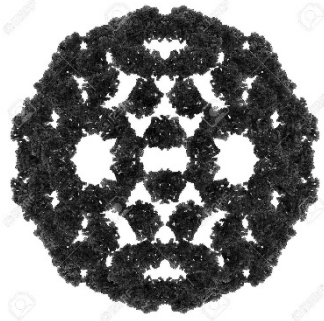


Tobacco mosaic virus

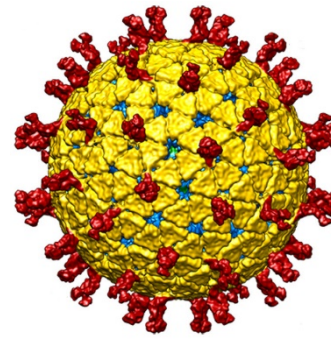


Parvovirus
(fifth disease)

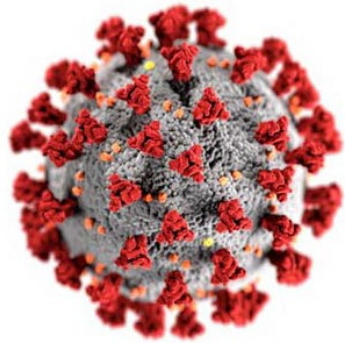
DNA VIRUS:
GROUP II: one strand



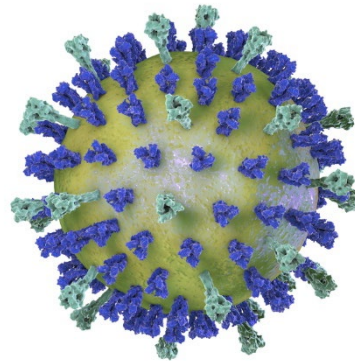
Bluetongue virus



Rotavirus
(gastroenteritis)



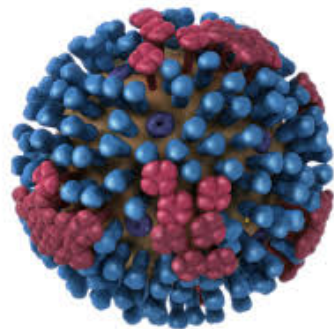
SARS-CoV-2



MERS



Yellow fever



Influenza virus



Ebola virus

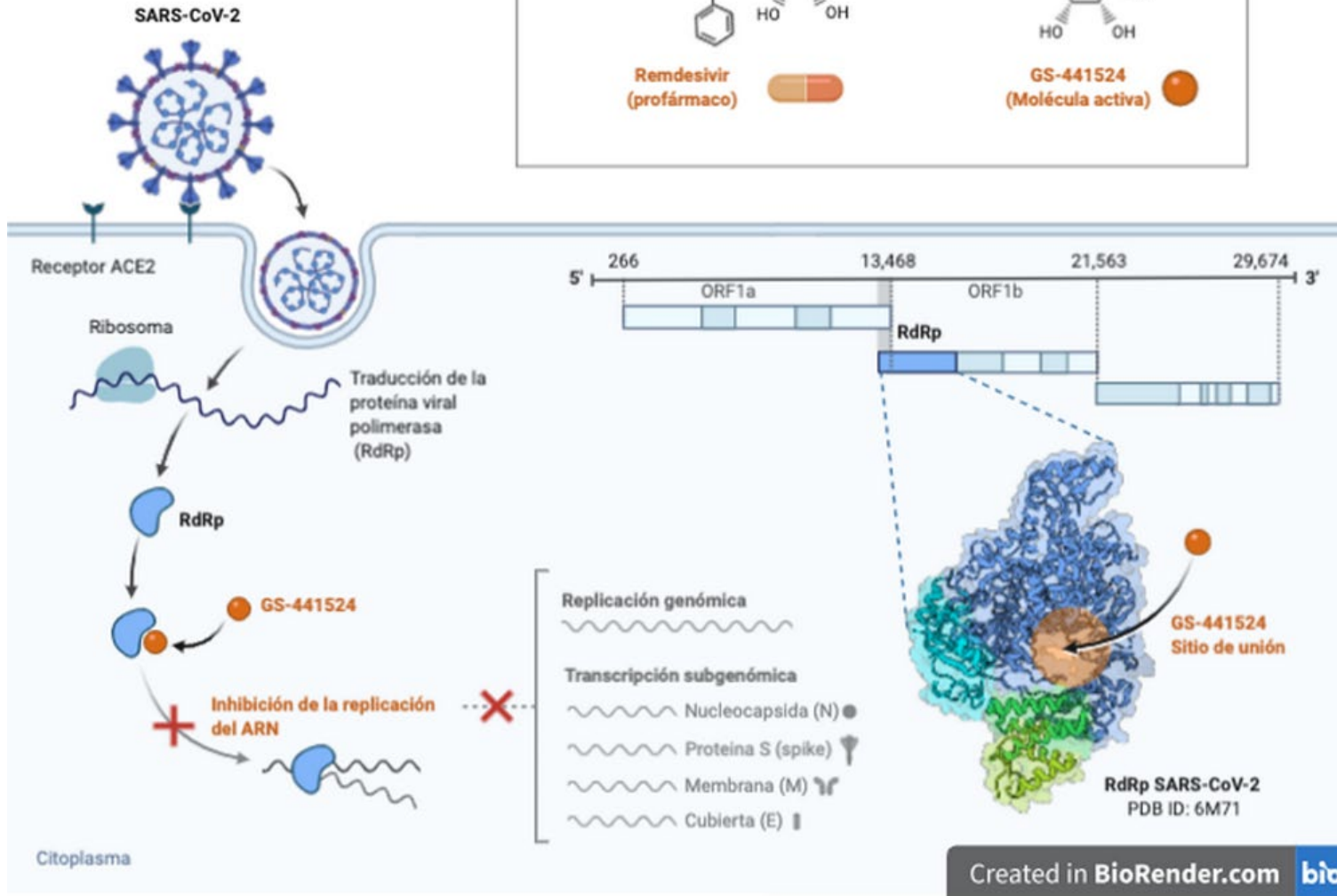
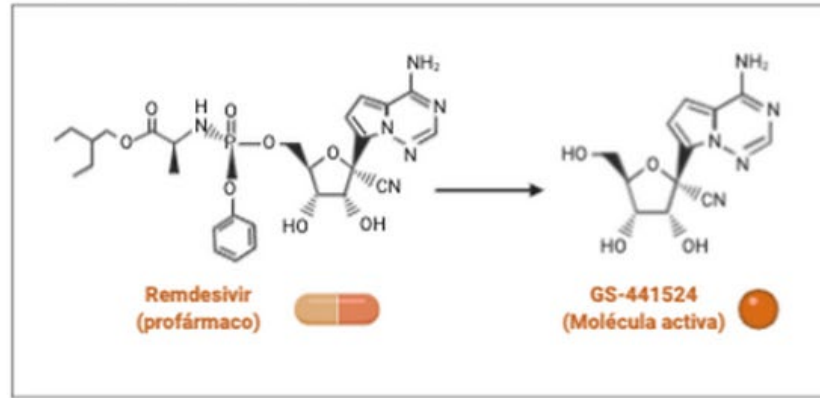
RNA VIRUS:
GROUP III: double strand

RNA VIRUS:
GROUP IV: one strand (+)

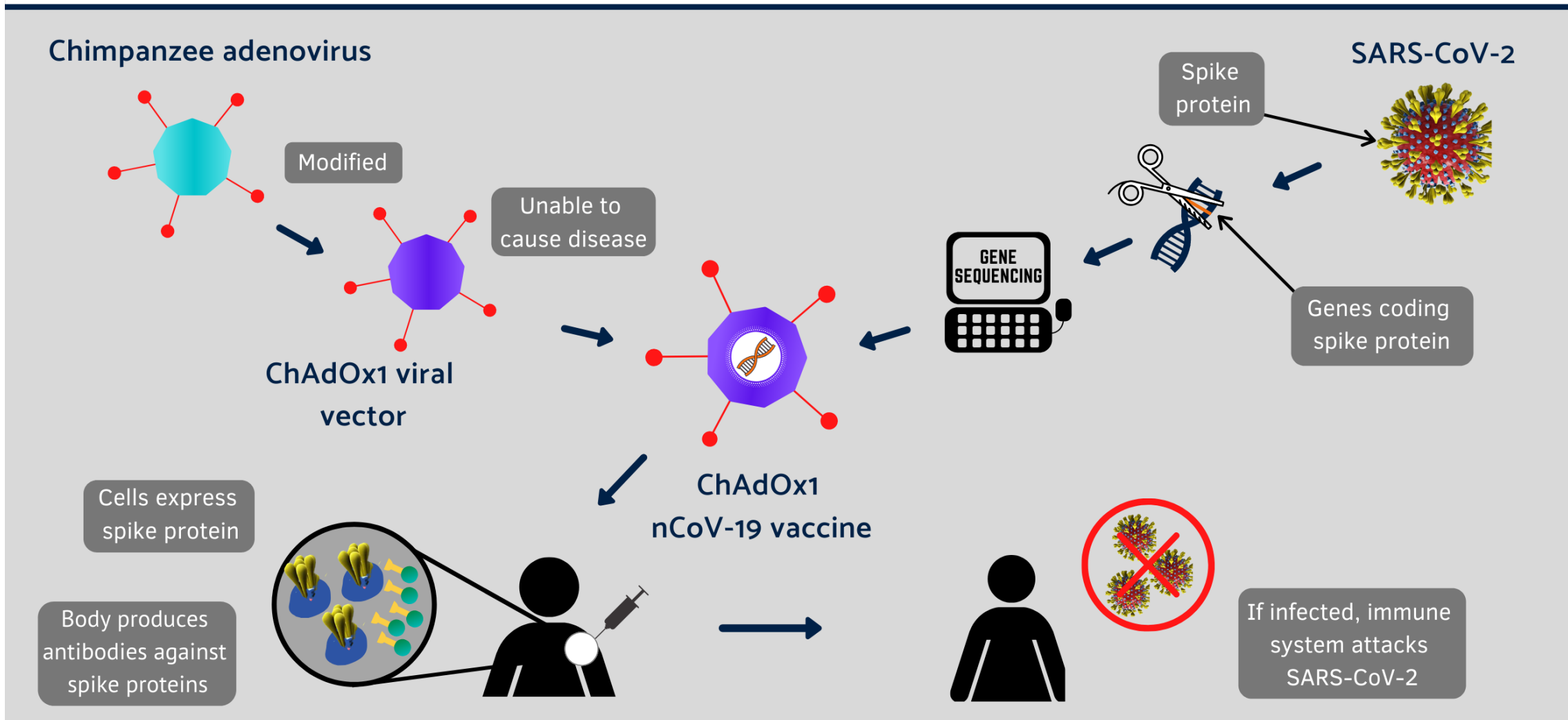
RNA VIRUS:
GROUP V: one strand (-)



Remdesivir



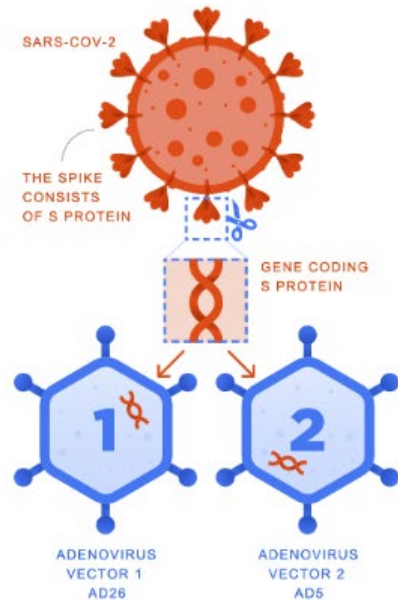
COVID-19 Oxford Vaccine Trial



Two-vector vaccine against coronavirus

Vector creation

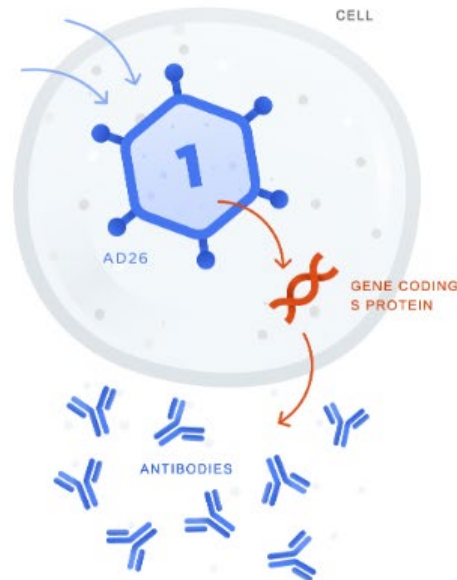
A **vector** is a virus that lacks a gene responsible for reproduction and is used to transport genetic material from another virus that is being vaccinated against into a cell. The **vector** does not pose any hazard to the body. The vaccine is based on an adenoviral vector which normally causes acute respiratory viral infections



A gene coding **S protein** of SARS-COV-2 spikes is inserted into each vector. The spikes form the "crown" from which the virus gets its name. The SARS-COV-2 virus uses spikes to get into a cell

First vaccination

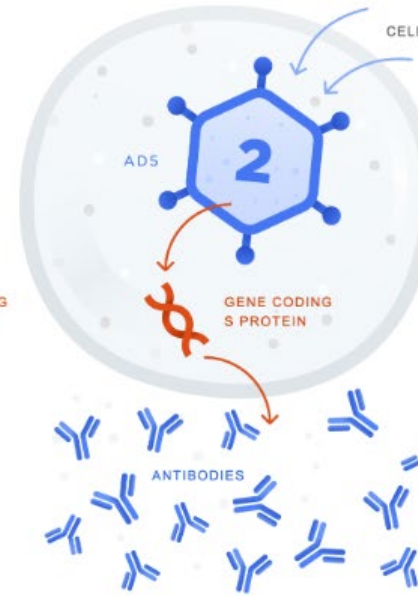
Vector with a gene coding **S protein** of coronavirus gets into a cell



The body synthesizes **S protein**, in response, the production of **immunity** begins

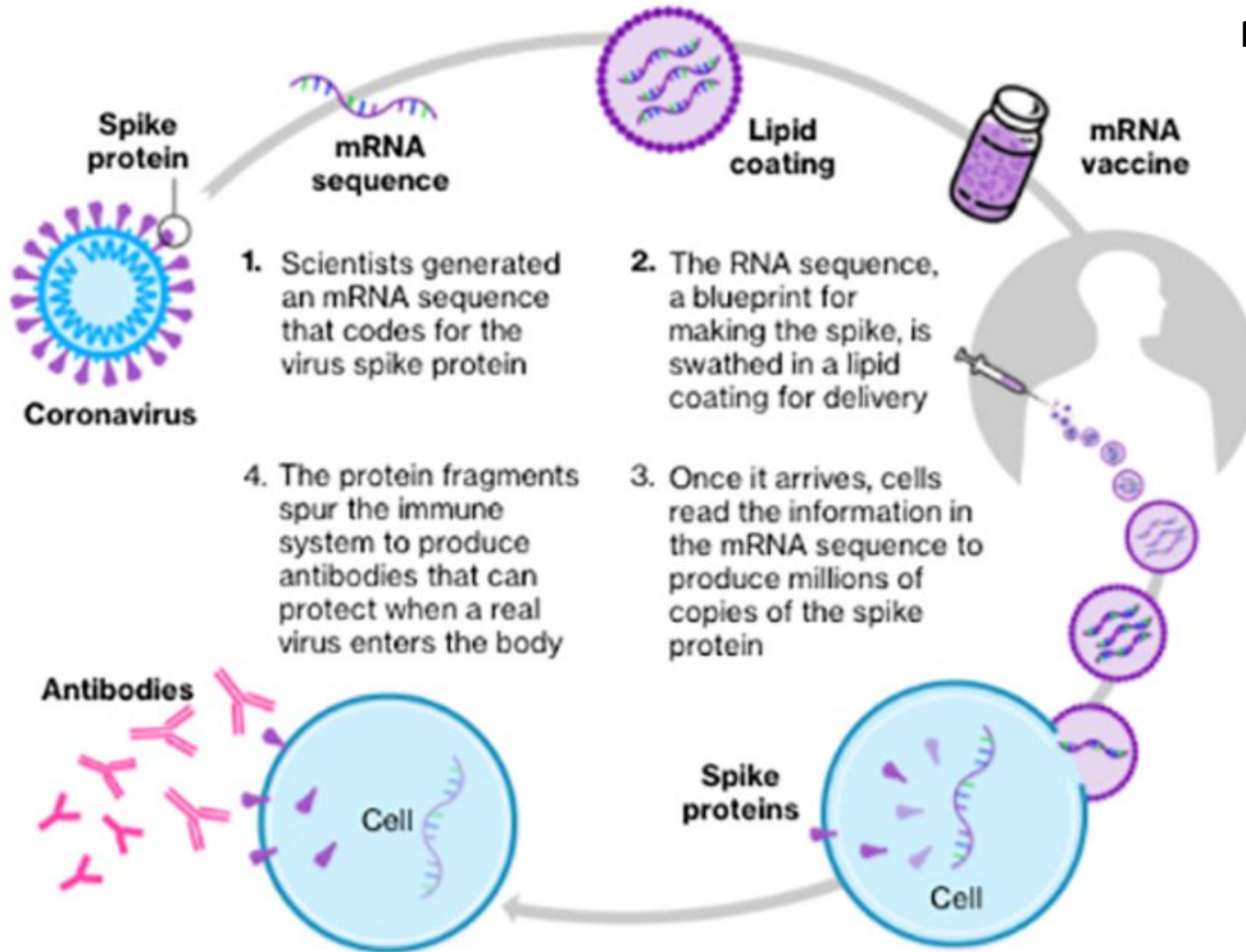
Second vaccination

Repeated vaccination takes place in 21 days

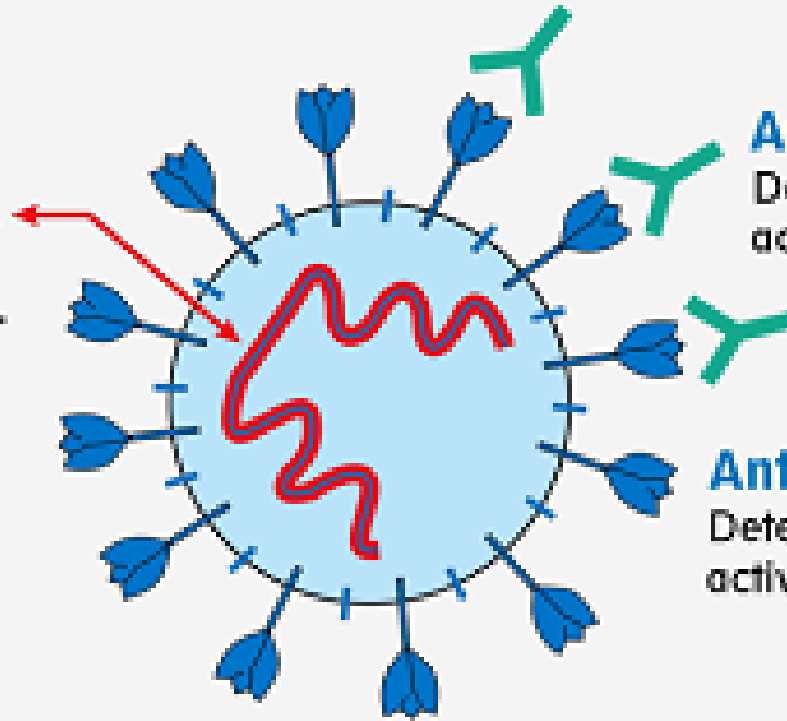


The vaccine based on another adenovirus vector unknown to the body boosts the immune response and provides for long-lasting immunity

The use of two vectors is a unique technology of the Gamaleya Center making the Russian vaccine different from other adenovirus vector-based vaccines being developed globally



RT-PCR
Detect RNA;
indicate active infection.



Antibody Test
Detect antibody; indicate
active or past infection.

Antigen Test
Detect antigen, indicate
active infection.

Features of RT-PCR:

- Gold standard diagnosis
- Turn-around time around 2 hours
- High equipment requirement

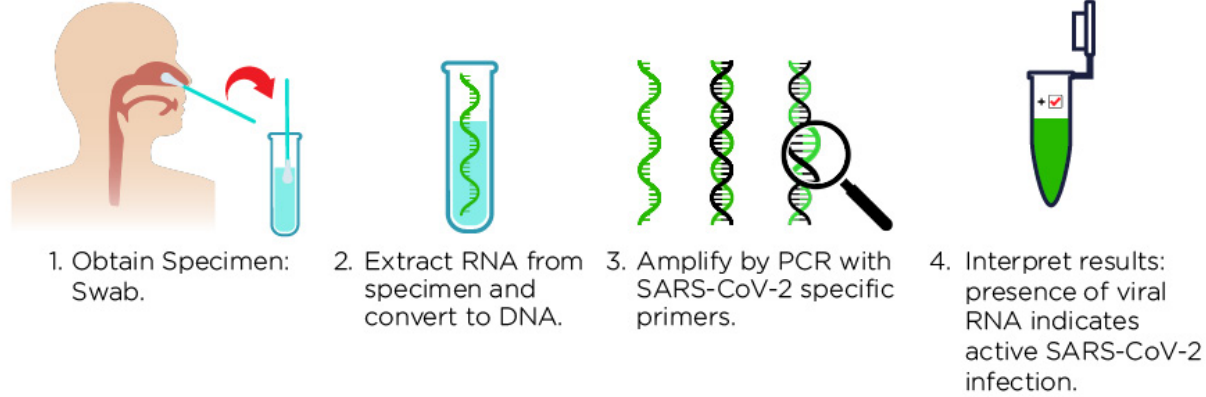
2019-nCoV

Features of Antigen & Antibody Test:

- Inexpensive cost
- Turn-around time around 15 mins
- Less equipment required

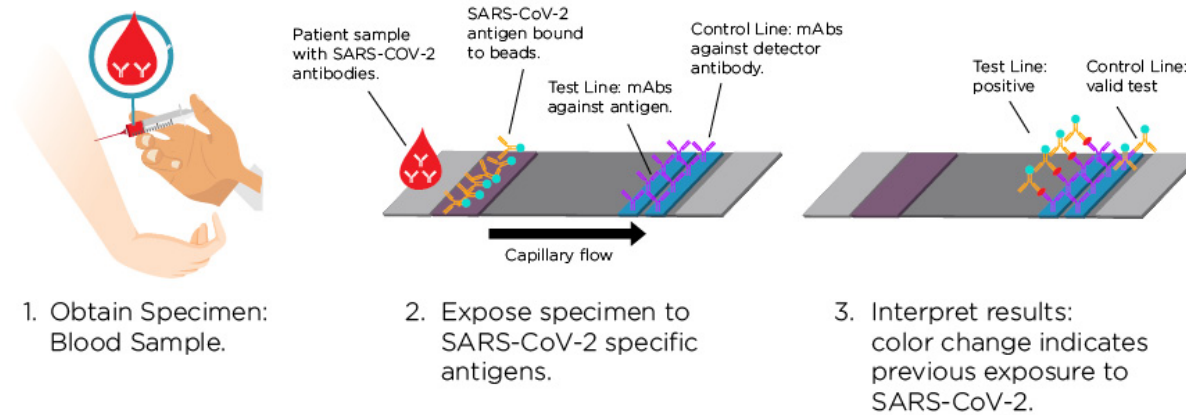
Molecular Tests (Nucleic Acid Detection)

Diagnose active SARS-CoV-2 infections



Antibody Tests (Serology)

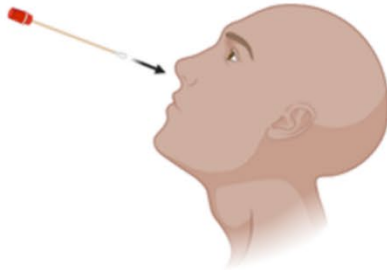
Detect immune response to SARS-CoV-2 exposure



COVID-19 Diagnostic Test using RT-qPCR technique

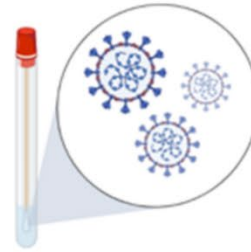
1 Nasopharyngeal swab <15 min

Cotton swab is inserted into nostril to absorb secretions.



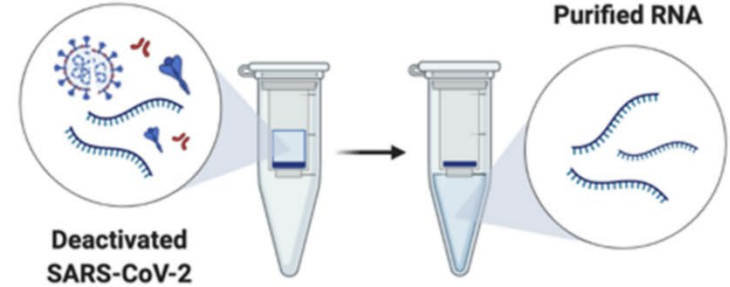
2 Collected specimen 0-72 h

Specimen is stored at 2-8°C for up to 72 hours or proceed for RNA extraction.



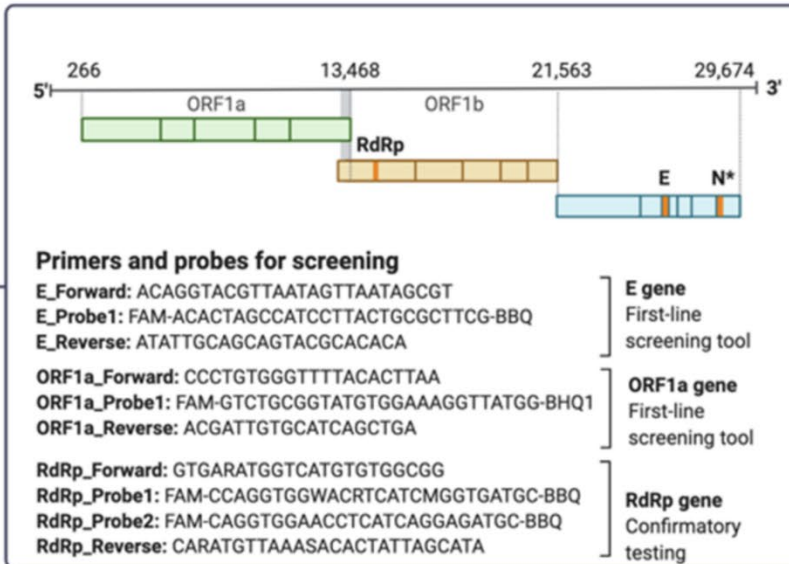
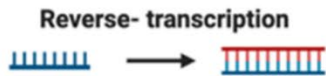
3 RNA extraction ~45 min

Purified RNA is extracted from deactivated virus.



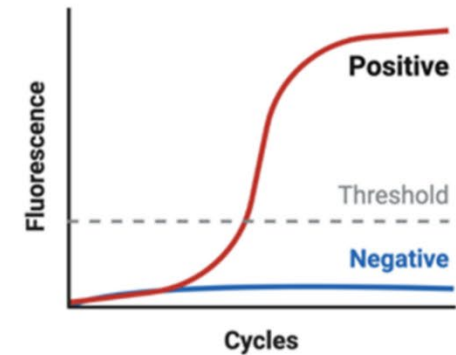
4 RT-qPCR ~1 h per primer set

Purified RNA is reverse transcribed to cDNA and amplified by qPCR.

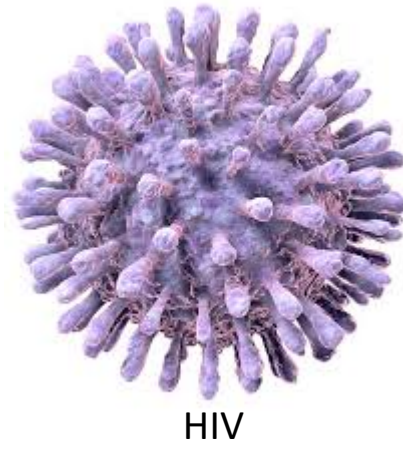
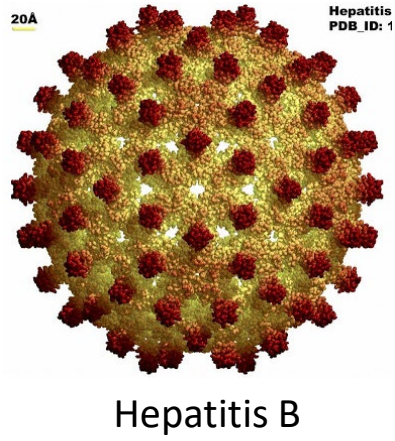


5 Test results real-time

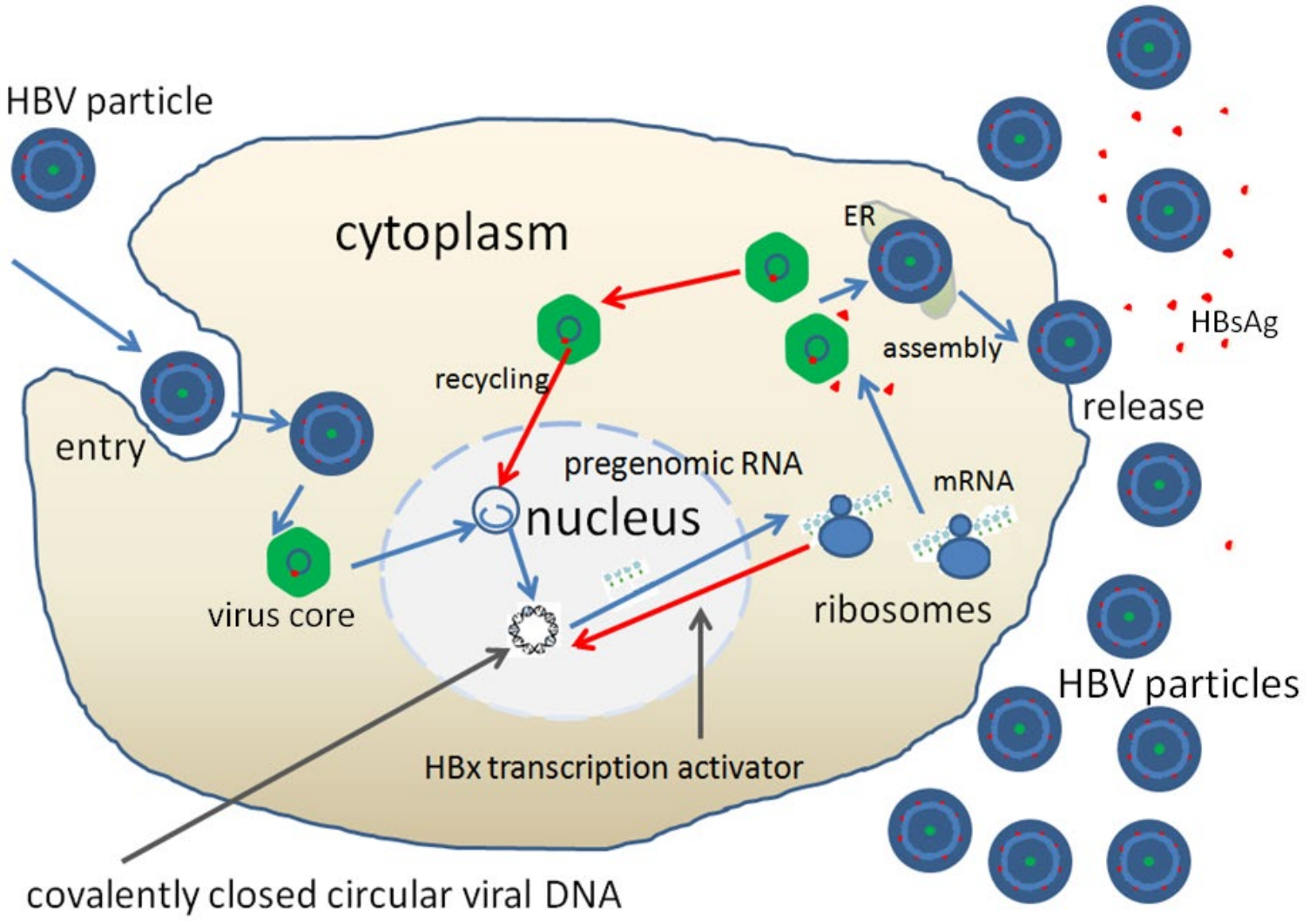
Positive SARS-CoV-2 patients cross the threshold line within 40.00 cycles (< 40.00 Ct).



DNA VIRUS OR RNA VIRUS WITH REVERSE TRANSCRIPTASE:
GROUP VI & VII

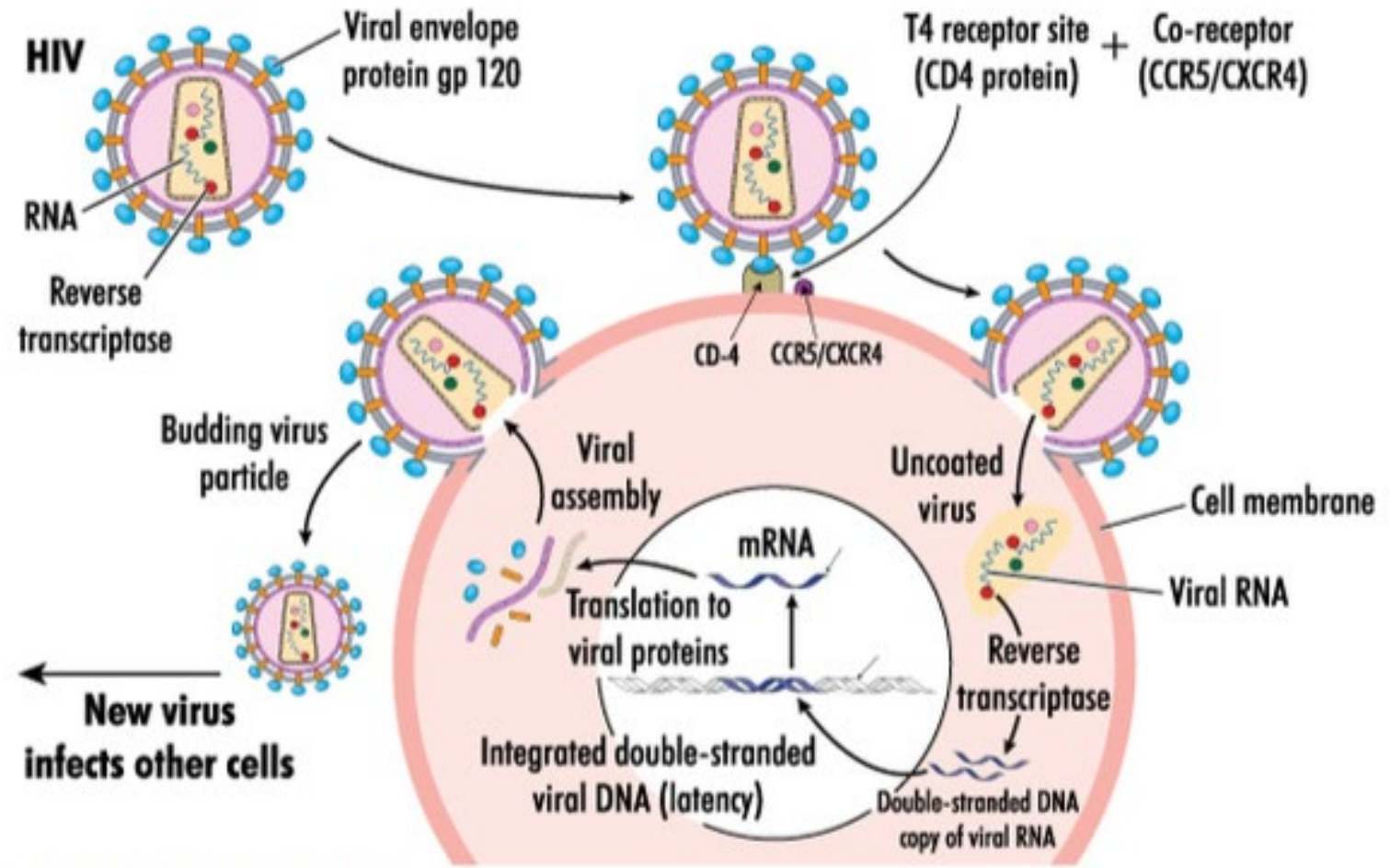


HBV CYCLE



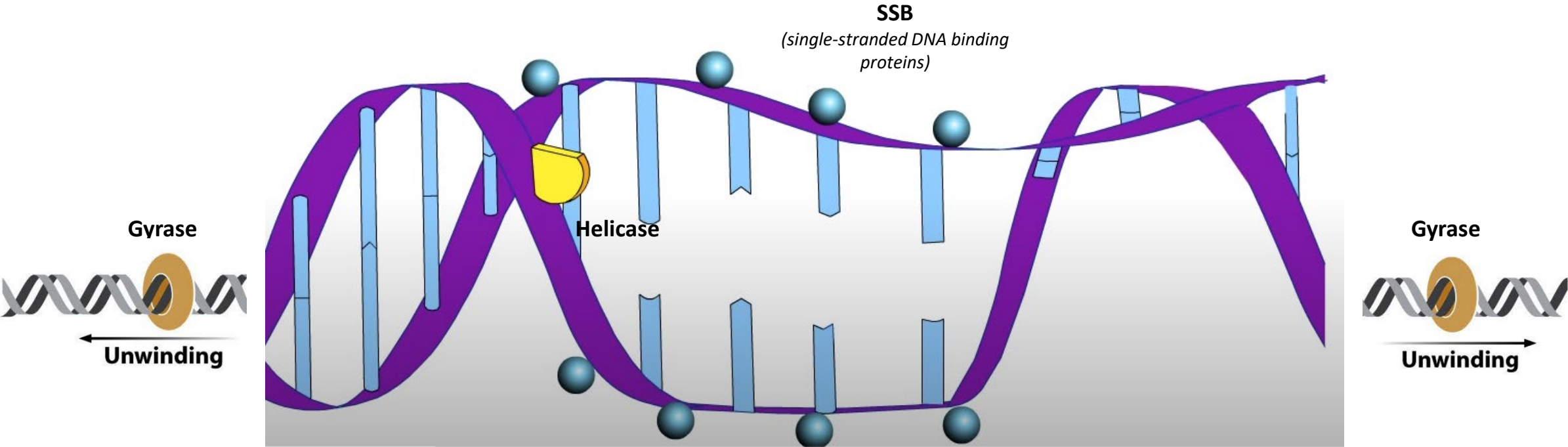
Source: Wikipedia

HIV CYCLE



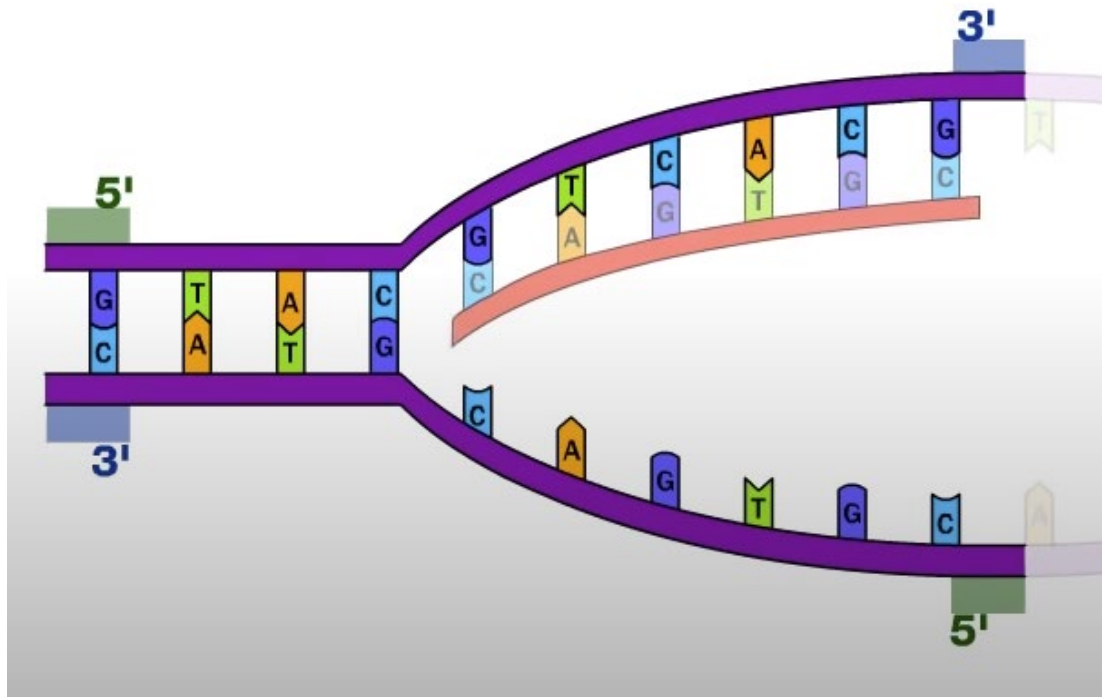
Credit: Blamb/shutterstock.com

DNA Replication



Source: <https://www.youtube.com/watch?v=YqjbmrQcyfM> & Panamericana

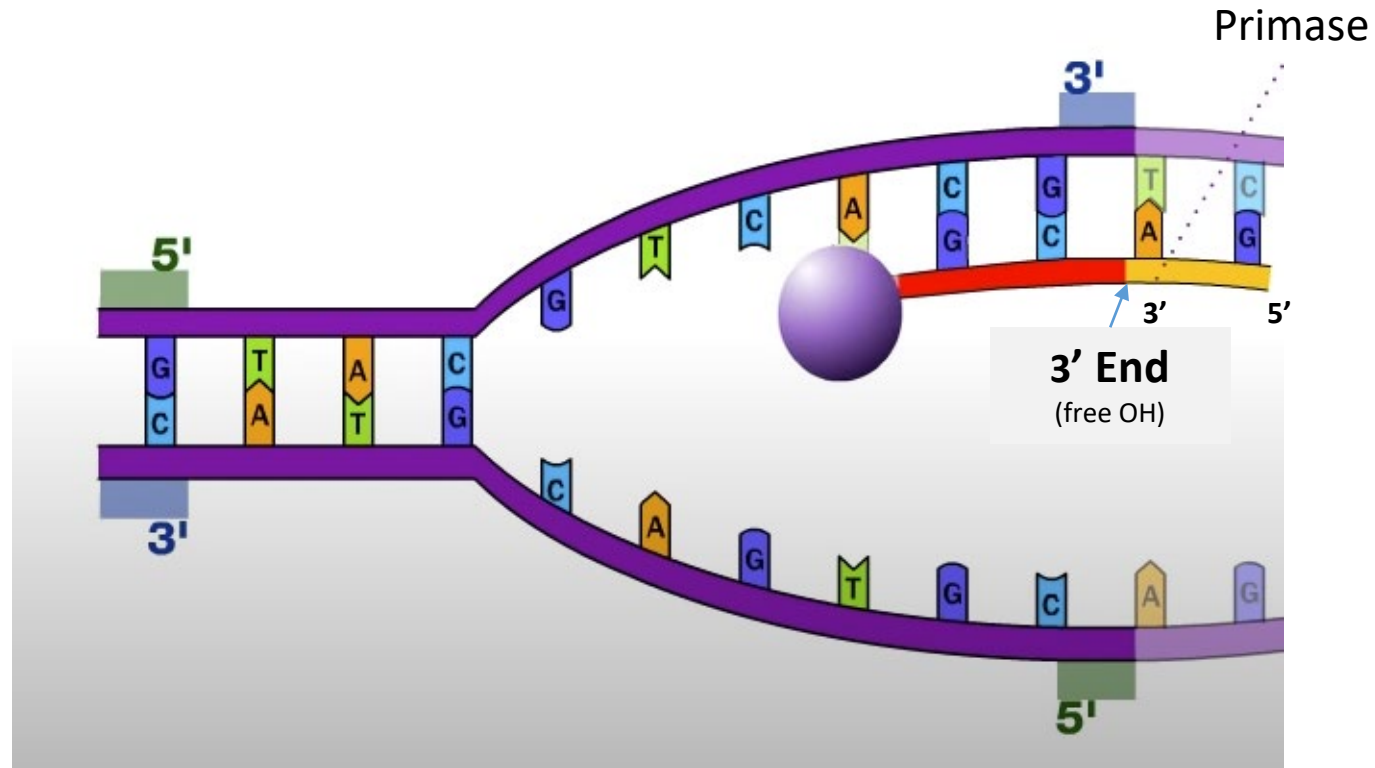
DNA Replication



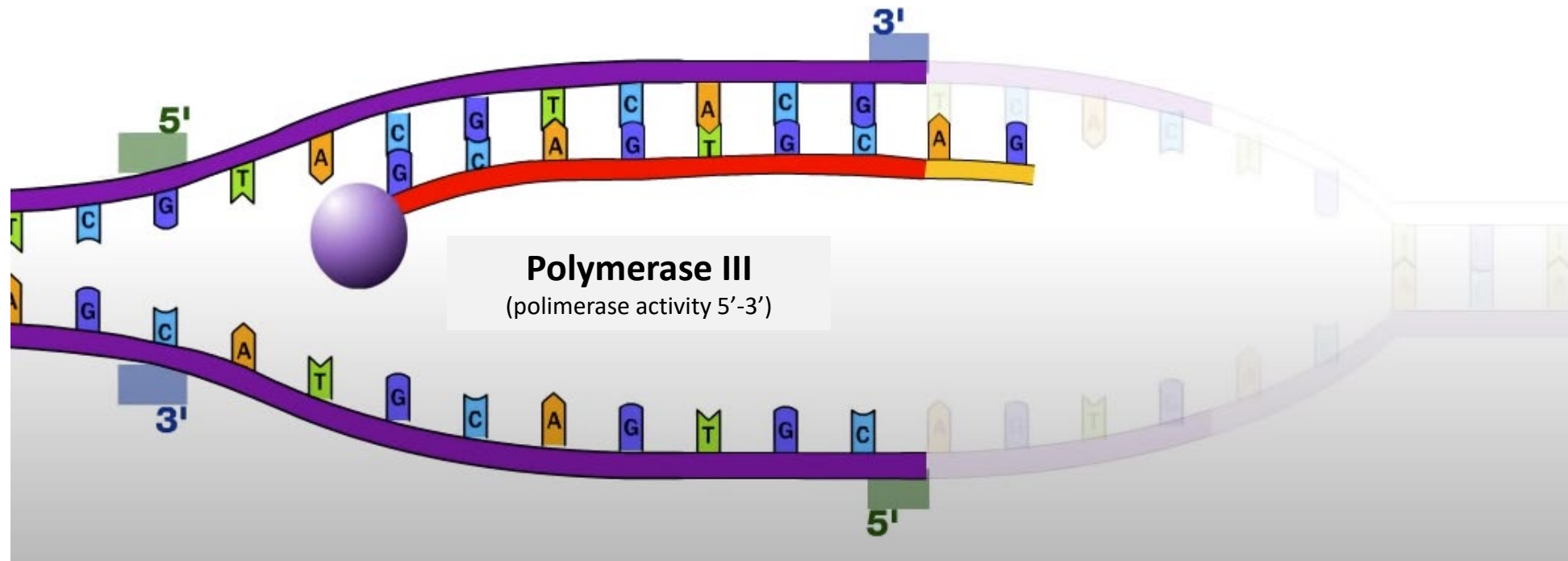
DNA POLYMERASE:

- Needs a double-stranded fragment (*primer*) with a free 3' end.
- Add nucleotides in **5'-3'** direction.

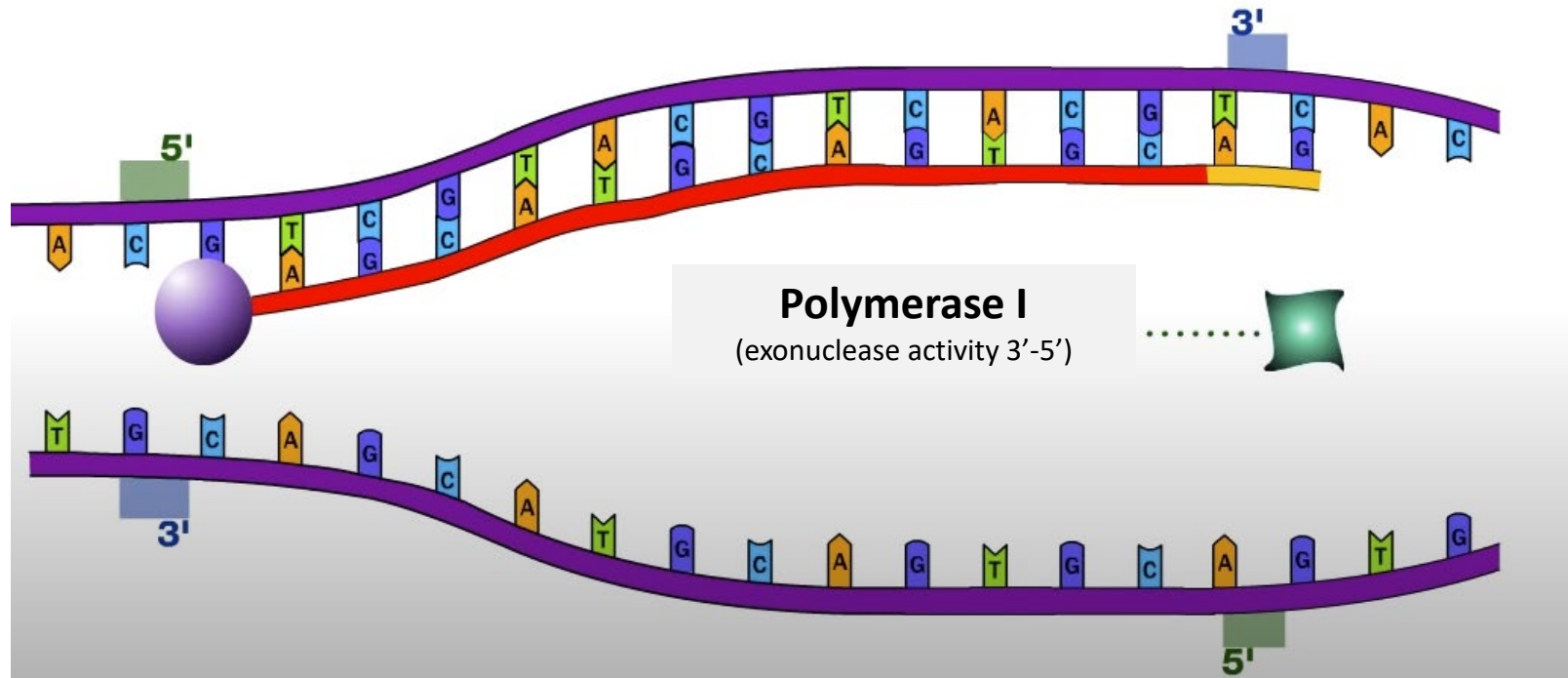
DNA Replication



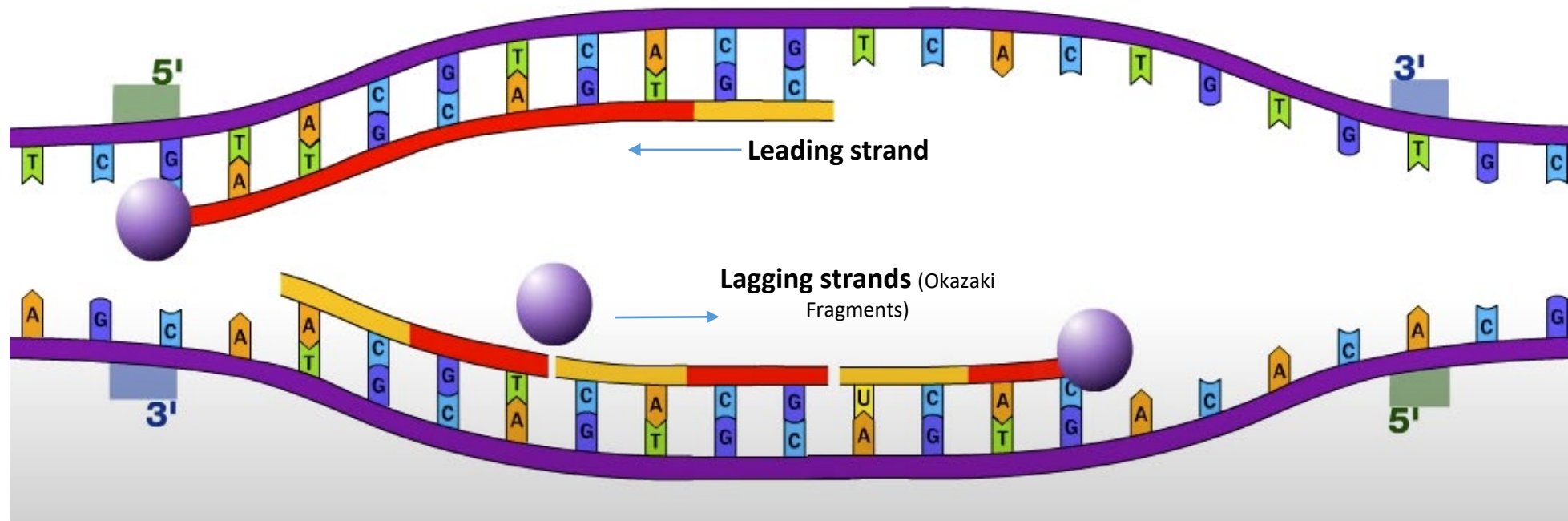
DNA Replication



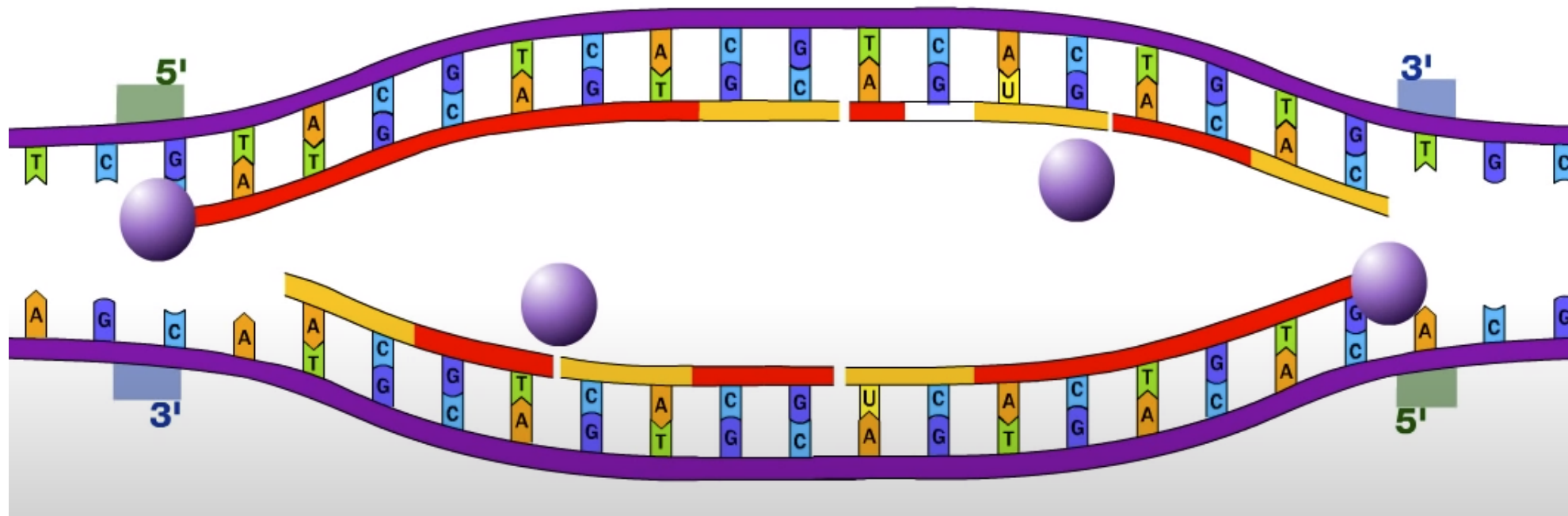
DNA Replication



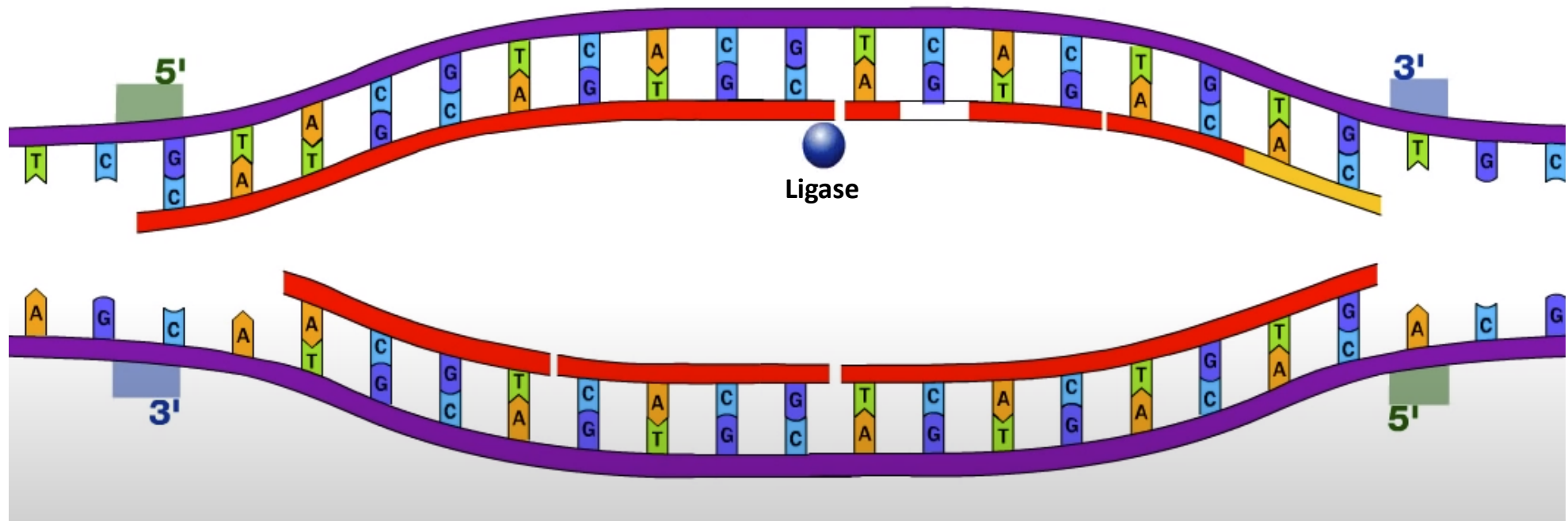
DNA Replication



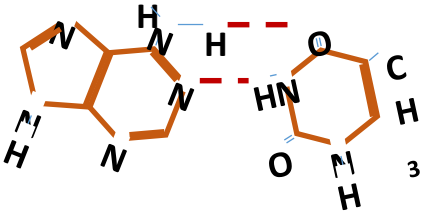
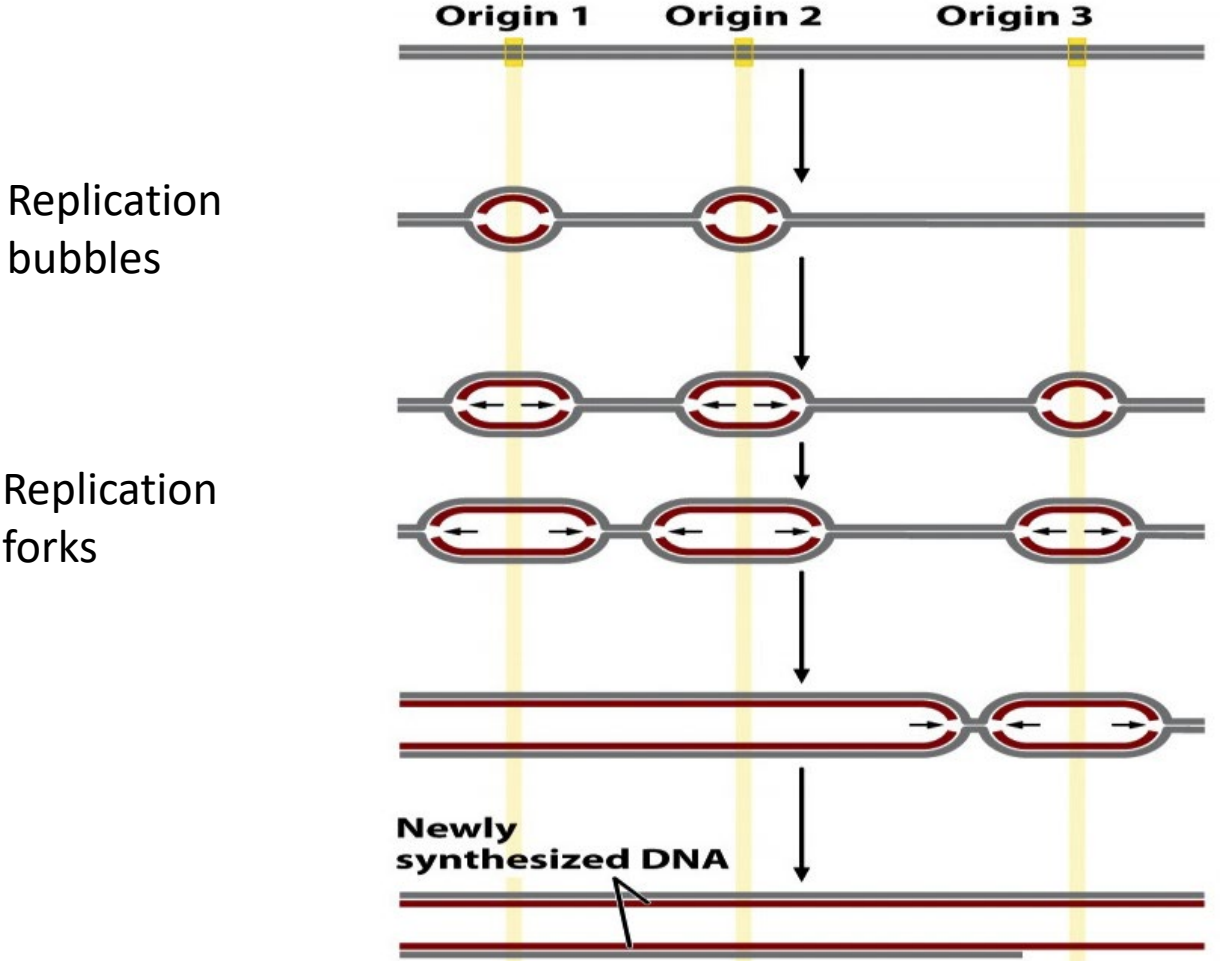
DNA Replication



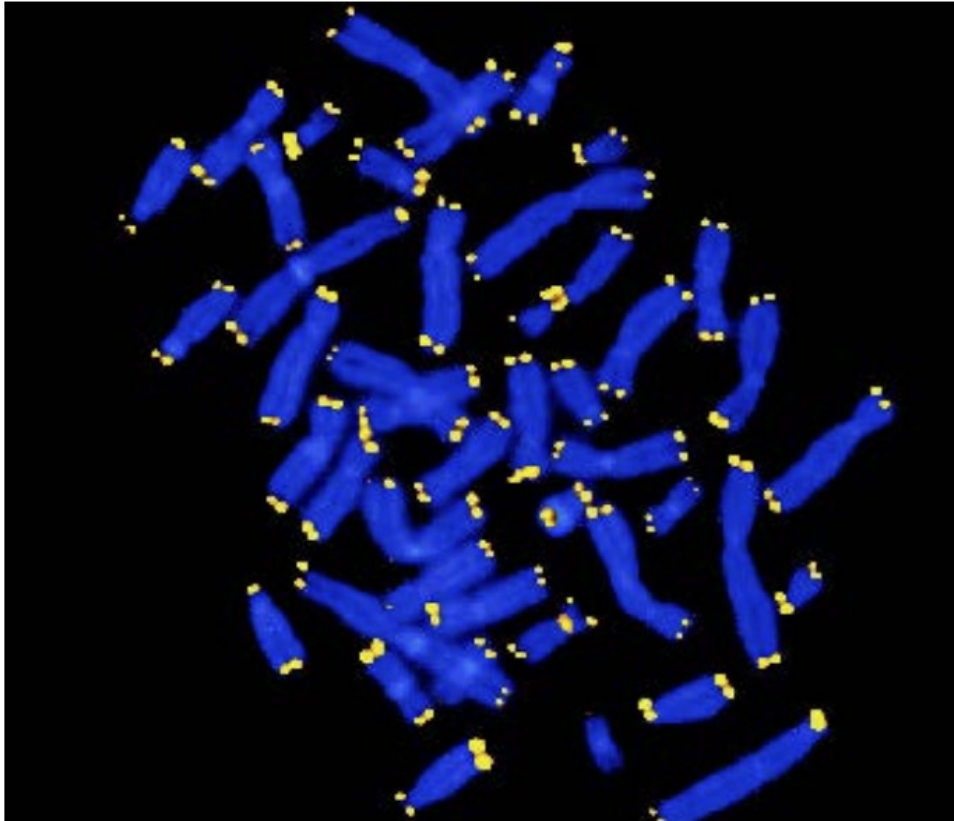
DNA Replication



DNA Replication

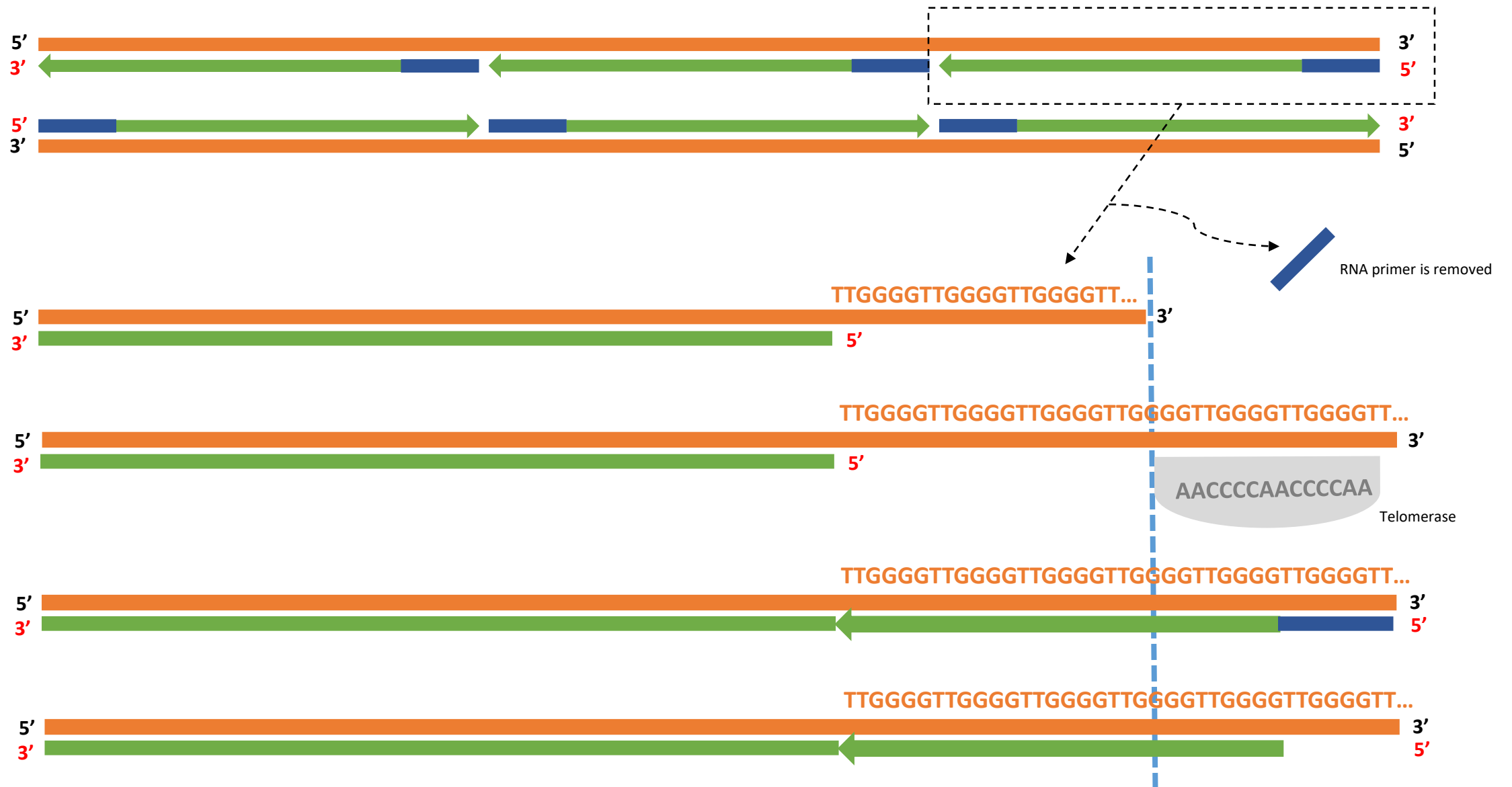


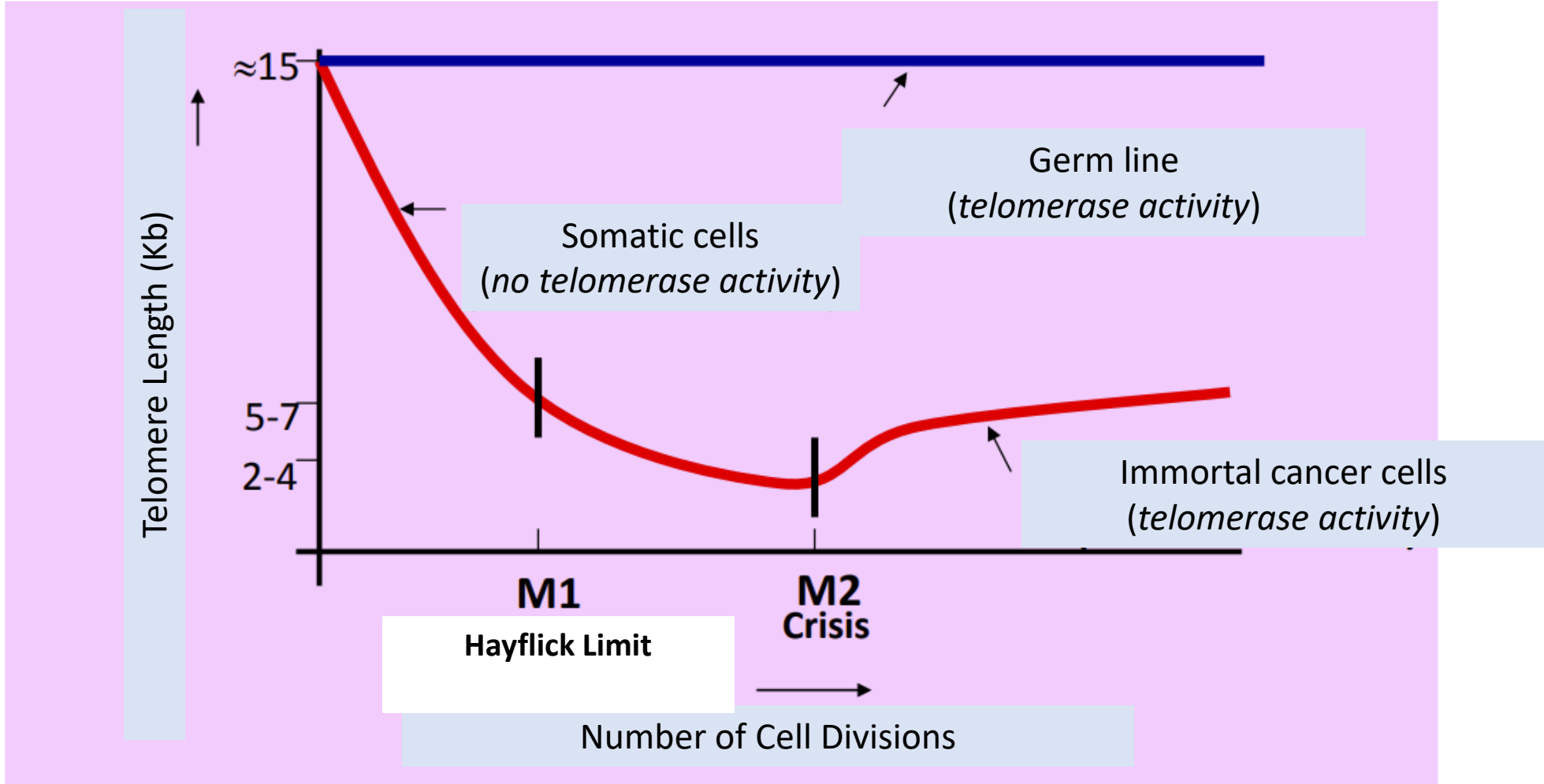
Replication of Telomeres



Species	Motif ^a (5'→3')
<i>Homo sapiens</i>	GGGTTA
<i>Saccharomyces cerevisiae</i>	G ₁₋₃ T
<i>Trypanosoma brucei</i>	GGGTTA
<i>Oxytricha fallax/nova</i>	GGGGTTTT
<i>Tetrahymena thermophila</i>	GGGGTT
<i>Stylonychia lemnae/pustulata</i>	GGGGTTTT

REPLICATION OF TELOMERES

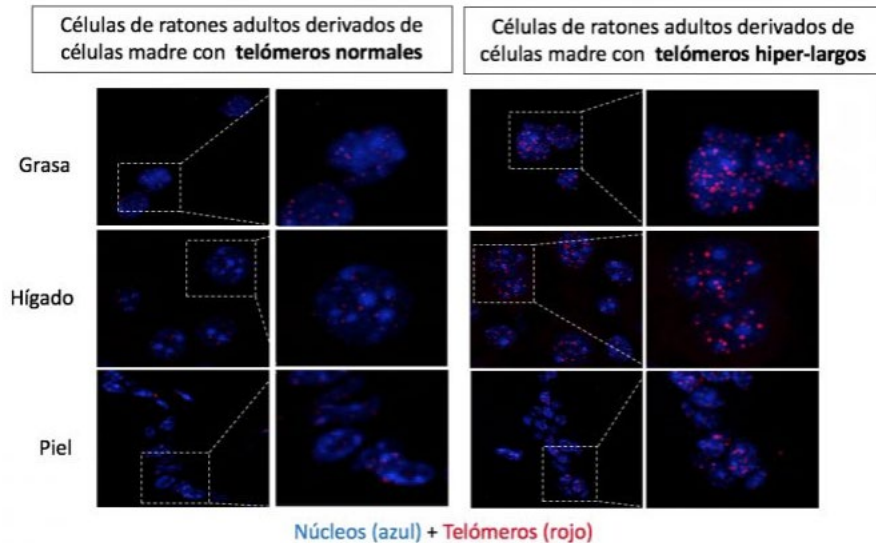




Nature Communications.
 Investigadores del CNIO
 consiguen los primeros
 ratones nacidos con
 telómeros hiperlargos y
 demuestran que es posible
 prolongar la vida sin ninguna
 modificación genética

17.10.2019

AYUDA A LA INVESTIGACIÓN



Portada > Salud > Biociencia

URGENTE

ANIMALES MÁS SALUDABLES EN SU VEJEZ

Científicos españoles crean ratones un 40% más longevos y resistentes al cáncer

Los animales viven más y también en mejores condiciones

Actualizado jueves 13/11/2008 18:00 (CET)



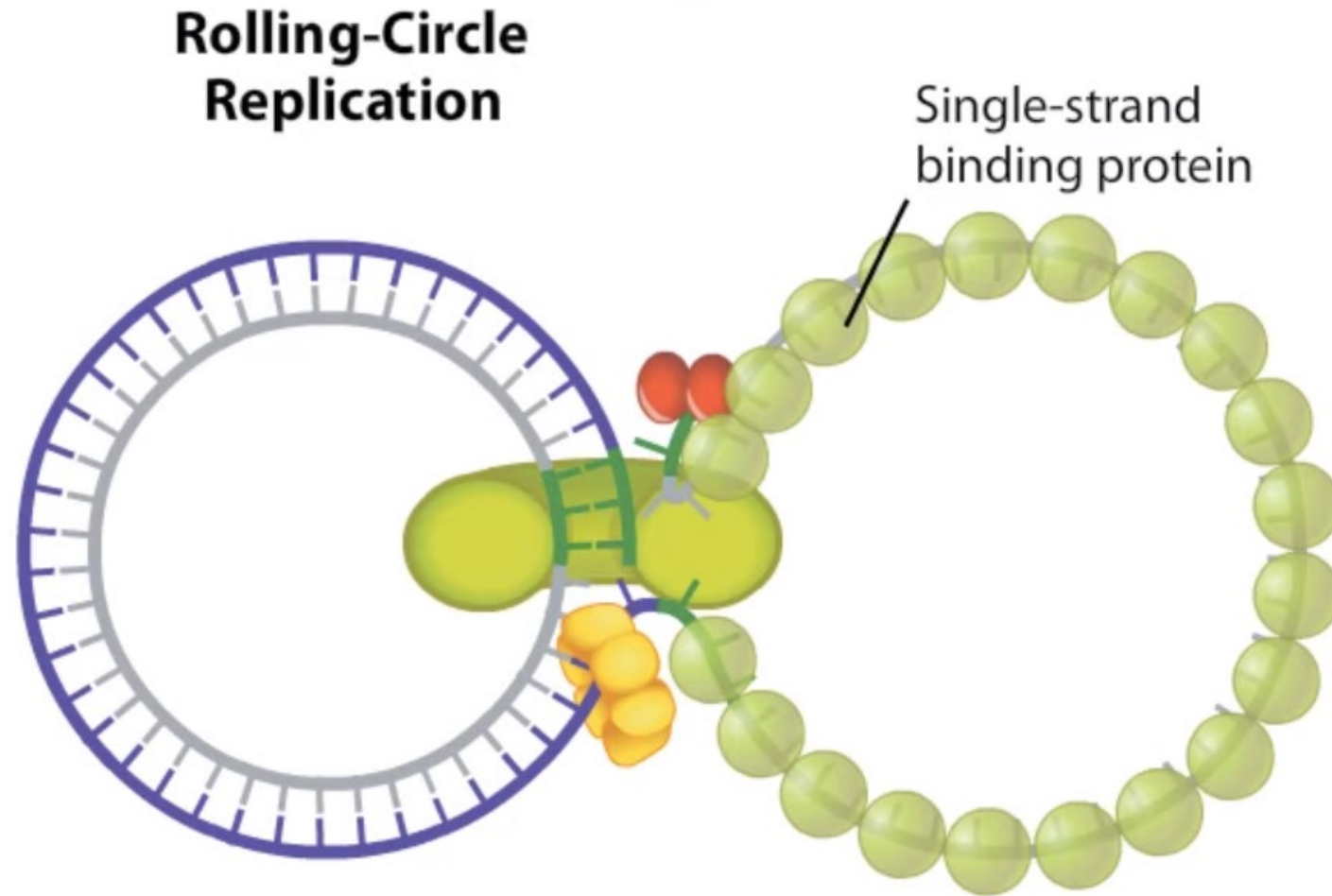
MARÍA VALERIO

MADRID.- Super Ratón, el personaje de dibujos animados de la factoría Terrytoons que volaba por las pantallas de televisión en la década de los ochenta con su capa roja, obtenía los poderes de una alimentación especial. En el laboratorio de María Blasco, en el Centro Nacional de Investigaciones Oncológicas (CNIO), los roedores obtienen sus 'superpoderes' de la telomerasa, una sustancia que protege a las células del envejecimiento y permite a los animales ser un 40% más longevos de lo normal.



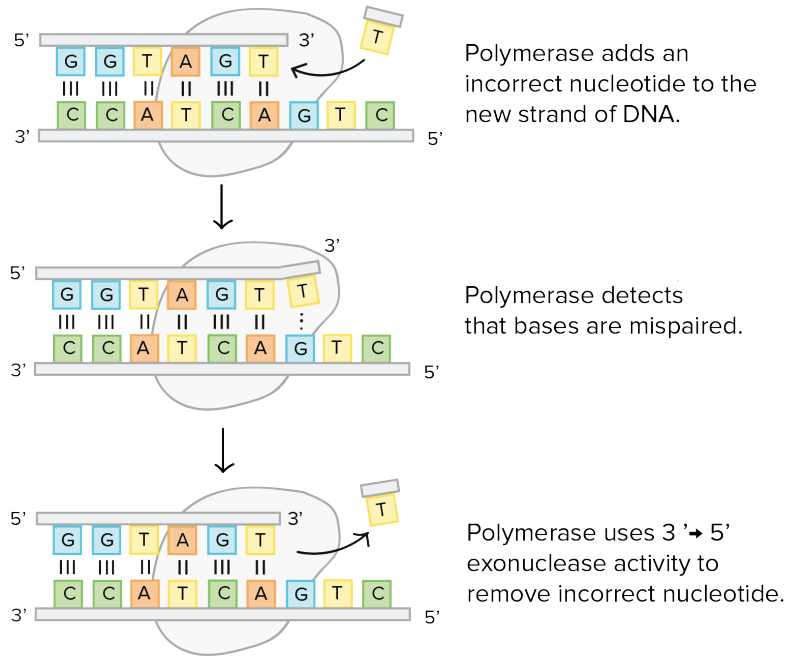
Para lograr la ecuación perfecta, el equipo de [Blasco](#) probó a modificar genéticamente a los ratones en una doble dirección: aumentando los niveles de telomerasa para que no envejeciesen e incrementando al mismo tiempo la presencia de varios genes supresores tumorales (p53, p16 y p19ARF) para protegerles del cáncer. Para ello, emplearon un modelo de ratón creado también en las mismas instalaciones del CNIO por el equipo de Manuel Serrano y que es 'inmune' al cáncer gracias a sus modificaciones genéticas; y sobre ese ejemplar aumentaron la proteína TERT, responsable de la regeneración de los telómeros.

DNA Replication in bacteria



Proofreading

Proof-reading



Mismatch repair

